Proposed upper limits for maximal LVEDD for white male is <60 mm and for black and mixed-race males <59 mm. For all females the cut-off is <52 mm.

Conclusions Black athletes of both sexes reveal the greatest wall thickness compared with mixed-race and white athletes. Conversely, white athletes display the largest cavity size in both sexes. Mixed-race athletes demonstrate electrocardiogram and echocardiogram features more similar to black athletes than white athletes. Information regarding the upper limits of normality in this age group is useful in enabling the differentiation between physiological adaptation and cardiomyopathy.

9 P62 DEFICIENCY PROTECTS AGAINST CEREBRAL ISCHAEMIA IN INSULIN RESISTANT MICE

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Introduction and aim Diabetic patients have poorer outcomes following ischaemic stroke. In rodents, similar findings are reported, with Type 1 and 2 diabetic rodents demonstrating increased infarct volume and worsened post-stroke recovery, which correlate with markers of autophagy overactivation post-ischaemia. p62 is a selective adaptor protein for autophagy, with mice lacking p62 (p62 KO) known to develop obesity-linked insulin resistance in adulthood and have suppressed autophagic flux. To ascertain whether p62 is associated with ischaemic stroke pathophysiology, we evaluated post-stroke infarct volume and functional recovery in wild type (WT) vs p62 KO mice.

Methods All procedures were performed in male 8–9 week old C57BL/6 mice in accordance with the ARRIVE guidelines and Home Office regulatory approval. Transient focal ischaemia was induced by middle cerebral artery occlusion (MCAo) for 1 hour followed by reperfusion for 72 hour. Adhesive removal and open field assays were conducted daily 3 days prior to and 72 hour post-stroke to assess sensorimotor and motor function. Mice were then euthanised and brains perfused-fixed to obtain 10 µm sections every 1 mm relative to bregma. Infarct volume and astrogliosis were quantified using nissl staining and GFAP immunofluorescence, respectively. Data were analysed by unpaired student's t-test or 2-way ANOVA and Bonferroni post-hoc test as appropriate.

Results Infarct volume assessed by nissl staining showed a significant ($p < 0.01$) reduction in p62 KO vs WT mice (16 ± 3 vs. $30 \pm 1\%$ brain volume, respectively). Proliferation and activation of astrocytes assessed by GFAP staining was found to be reduced in the peri-infarct border in p62 KO vs WT mice ($p < 0.05$). Furthermore, p62 KO showed marked improvements in post-reperfusion functional recovery. Reduced motor function was observed in WT mice, with a ~70% reduction in max speed and total displacement vs baseline observed 72 hour post-reperfusion ($p < 0.01$). No such impairments were reported in p62 KO mice. Similarly, sensorimotor function was impaired in WT mice post-reperfusion, with an increase in adhesive removal time in the affected (227 ± 32 vs 29 ± 9 ms at baseline, $p < 0.001$) and unaffected (73 ± 23 vs 16 ± 3 ms at baseline, $p < 0.05$) paw. p62 KO mice were protected, with adhesive removal times in the affected (51 ± 44 vs 28 ± 7 ms at baseline, $p < 0.05$) and unaffected (61 ± 39 vs baseline 28 ± 7 ms

at baseline) paws substantially improved when compared to WT mice at 72 hour.

Conclusions Our findings show that p62 KO mice are protected from cerebral ischaemia. This protection persists despite p62 KO mice being glucose intolerant, suggesting that p62 may be a meaningful therapeutic target even in comorbid diabetic populations. Further studies are needed to ascertain whether short-term functional recovery afforded by p62 deficiency persists long-term.

10 EVIDENCE THAT ALTERED REDOX STATUS RESULTS IN KCA3.1 CHANNEL REDUCED ENDOTHELIAL CELL SURFACE EXPRESSION

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Introduction Endothelial derived hyperpolarization (EDH) is an important path to vasodilatation especially in small arteries, and is more important in women and the elderly. Our previous work has shown that this becomes impaired in mice that had experienced a high fat diet during gestation up to weaning, with concomitant reduction in endothelial surface KCa3.1 expression.¹ We have tested the idea that altered redox status affects the location of KCa3.1 in human dermal microvascular endothelial cells (HDMECs).

Methods HDMECs were obtained from PromoCell (Heidelberg, Germany) and cultured in MV2 growth medium containing 1% penicillin-streptomycin in 5% O₂, 5% CO₂ at 37°C to passage 6 and 7 in the SCI-tive Physiological Oxygen Workstation from Baker Ruskinn (Maine, USA) for at least 5 days before experiments. The cells were plated on fibronectin (10 µg.ml⁻¹) coated 8 well micro-slides (Ibidi, Martinsried, Germany) and incubated for 48 hour, treated with PBS or H₂O₂ 100 µM in PBS for 10 min., and fixed with 4% formalin. Immunofluorescence was performed by incubating with anti-KCa3.1 mouse monoclonal antibody (AL-051 Alomone, Israel) at 4°C overnight, with half of the wells being treated with 2% Triton-X to permeabilize the cells. The Alexa 488 labelled anti-mouse antibody for 1 hour and Texas Red Lycopersicon Esculentum Lectin (to stain the cell surface) for 20 min, were applied to the cells at room temperature and finally the DAPI nuclear stain was applied for 3 min. The cells were examined using a x60 1.3 NA oil-immersion objective and images captured via a Hamamatsu RG3 camera into a computer. Using ImageHopper software and computer-driven focussing device (Prior Instruments, UK) we took 10 serial images of KCa3.1 at 1.0 µm intervals, which were integrated using Image J. These images were analysed for the fractional area of an endothelial cell occupied by KCa3.1, defined as bright spots between 1 to 4 µm diameter in the relevant fluorescence channel.

Results The density of channels on the surface, defined as the integrated image of non-permeabilized KCa3.1 channels, was $9.5 \pm 0.68 \mu\text{m}^{-2} \times 10^3$ (mean $\pm$ sem), while in the permeabilized cells the density within the whole cell area was $13.9 \pm 1.89 \mu\text{m}^{-2} \times 10^3$ ($p < 0.05$, 't' test, 4 experiments with 20 cells per experimental group) indicating that about 70% of the channels normally reside on the cell surface. Treatment with H₂O₂ had little effect on the total cell density (reduced