Light chain deposition disease presenting with hepatomegaly: an association with amyloid-like fibrils


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Summary: We report an unusual case of lambda light chain deposits without overt plasma cell dyscrasia. The clinical presentation was hepatomegaly without biochemical sign of renal involvement. Portal hypertension, spontaneous rupture of the spleen and fracture of the 12th thoracic vertebra occurred during the course of the disease. Ultrastructural studies showed that lambda light chain deposits were associated with amyloid-like deposits. This case suggests that light chain deposition disease and amyloidosis could be two expressions of the same disease.

Introduction

Since Randall et al. described the light chain deposition disease (LCDD) which is a systemic disease with constant and predominant renal involvement, numerous cases of glomerulonephritis with light chain deposits have been reported. We report here a case of LCDD unusual in that the clinical presentation was hepatomegaly without biochemical evidence of renal involvement and ultrastructural studies showed that light chain deposits were associated with amyloid-like deposits.

Case report

A 63 year old woman was referred in March 1984 for epigastric pain and a 10kg weight loss. On physical examination a 15cm large, regular and firm hepatomegaly was palpated. Total serum bilirubin was 44 µmol/l, serum alkaline phosphatase 469 IU/l (normal 30–80), serum aspartate aminotransferase 47 IU/l (normal 5–30). Prothrombin time (expressed as per cent of normal) was 75%, serum albumin 35 g/l and serum creatinine 71 µmol/l. Proteinuria and microscopic haematuria were absent. An abdominal ultrasound examination showed homogeneous hepatomegaly; the kidneys appeared normal. At laparoscopy the liver was regular, pale and enlarged. A liver biopsy was performed and LCDD was diagnosed on the presence of an amorphous material negative for amyloid stainings (Congo red, thioflavine T) but positive for anti-lambda light chain staining (Figures 1 and 2). It was distributed along the spaces of Disse and the centrilobular and portal areas and outlined the sinusoids and laminated the hepatic plates (Figure 1). There was no peliosis. Bone marrow aspirate, serum and urine immunoelectrophoresis were normal. A bone marrow biopsy was performed (see pathological findings). Skeletal X-rays just showed an axial demineralization. The patient received cyclophosphamide, melphalan and prednisolone from April to October 1984. In June 1984, spontaneous rupture of the spleen occurred requiring splenectomy. Bone marrow aspirate, serum and urine immunoelectrophoresis were normal. The spleen contained multiple cystic nodules 1–15 mm in diameter filled with red cells and fibrin and surrounded by amorphous material.

The patient was again hospitalized in November 1984 because of sudden thoracic pain with weakness of the legs. Skeletal X-rays showed diffuse demineralization with collapse of the 12th thoracic vertebra; no other lesion could be detected. Bone marrow aspirate, urine and serum immunoelectrophoresis were still normal. Laminctomy and a stabilization of the thoracolumbar spine by a plate

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Figure 1 Liver biopsy shows an amorphous material along the spaces of Disse, which laminates the hepatocytic plates. Trichrome stain, magnification ×672.

Figure 2 Immunofluorescence study of the liver: anti-lambda light chain gives a positive staining along the sinusoidal spaces. Magnification ×420.

were performed. The vertebral body was destroyed by a diffuse amorphous material which extended into the paravertebral muscles and ligaments.

She was readmitted on 24 December 1984 for encephalopathy. Physical examination showed asterixis, jaundice, ascites and oedema of the legs. Ascitic protein concentration was 16 g/l. Serum bilirubin was 398 µmol/l and serum creatinine 153 µmol/l. There was still no proteinuria. Bone marrow aspirate, serum and urine immunoelectrophoresis were normal. Ultrasound examination showed recanalization of the umbilical vein. The patient died in coma one month after admission. An autopsy was performed.

In 15% of the glomeruli there was moderate mesangial proliferation, and amorphous material was visible in the interstitium and in some capillary and tubular walls. In the liver the sinusoidal deposits appeared more abundant than in March 1984, and hepatocytic plates were considerably laminated.

In the liver, bone marrow, spleen and thoracic vertebrae, the deposits were Congo red-negative and showed no birefringence under cross-polarization. Thioflavine T staining was negative. Known amyloid liver sections were used as controls.

Direct immunofluorescence was performed on hepatic and renal frozen specimens from biopsy
(liver) and autopsy (liver and kidneys). Commercial fluoresceinated monospecific antisera against lambda and kappa immunoglobulin chains, C3, Clq, C4, fibrin and albumin were used (Behringwerke and Sebia Dako). Positive and negative kidney specimens served as controls. Only anti-lambda light chain gave a positive staining. In the liver, a strong staining was observed along the sinusoidal spaces and in the vascular walls, in the interstitium and around the biliary ductules of the portal areas (Figure 2). In 75% of the interstitium and around the biliary ductules of the portal areas (Figure 2). In 75% of the glomeruli, a bright and coarse staining was observed in the mesangial areas, along the glomerular and tubular basement membrane and the capillary walls. On bone marrow and thoracic vertebral body sections, positive stainings for IgG, IgA, IgM, lambda and kappa chain were observed in plasma cells showing a polyclonal immunoglobulin secretion.

Electron microscopy was performed on material from biopsy (liver) and autopsy (liver and kidneys). In both liver and renal specimens, ultrastructural study showed the presence of finely granular electron-dense basement-like material which appeared nearly black. In the liver, this material was essentially located in enlarged Disse spaces which were filled by irregular clumps of granular substance (Figure 3). The deposit was also observed in the portal areas, along the collagen fibrils, the basement membrane of biliary ductules and in the vascular walls. The glomeruli showed prominent nodular mesangial areas. There was striking diffuse infiltration of both mesangial matrix and peripheral capillary basement membranes by the dense granular deposit (Figures 4 and 5). Some granular deposits were also observed in tubular basement membranes, interstitium and blood vessels. In some glomeruli, there was a fibrillar deposit associated with granular deposits (Figures 4 and 5). The fibrillar material was similar to amyloid fibrils: it was formed by criss-crossing, 9 to 10 nm thick, randomly oriented bundles of fibrils (Figure 5).

**Discussion**

In our patient the diagnosis of LCDD was based on the presence in liver, spleen, bone and kidneys of an amorphous material which reacted with an anti-lambda light chain antiserum and had a granular aspect at electron microscopy. However, this case of LCDD was unusual because of its clinical presentation and the association of amyloid-like and light chain deposits.

Renal involvement is observed in all cases of LCDD and almost always reveals the disease. In our patient, LCDD was discovered by hepatomegaly when there was no biochemical sign of renal involvement. To our knowledge, only 2 other cases of LCDD have presented with hepatomegaly but proteinuria was present at the time of diagnosis. In our patient post-mortem microscopic study showed some renal deposits, but renal failure only appeared 3 weeks before death. Portal hypertension occurred during the course of the disease as attested by recanalization of the umbilical vein and the presence of ascites with low protein concentration. Portal hypertension is uncommon in LCDD, reported in only 2 other cases. It might be explained by the presence of abundant deposits in

![Figure 3](http://pmj.bmj.com/) Liver biopsy – an electron photomicrograph shows the presence of abundant granular dense deposits in Disse space (arrow). Magnification ×6790.
the Disse spaces which caused sinusoidal obstruction of the portal venous system.

In our case, two unusual complications appeared during the course of the disease: spontaneous rupture of the spleen and fracture of the 12th thoracic vertebra producing spinal cord compression. Such complications have not been reported previously, even though spleen and bone involvement have been described.1,2,5 The appearance of peliosis in the spleen was the same as that observed in the liver among some patients with LCDD.2,3,5

Another peculiarity of our patient was the ultrastructural aspect: fibrillar amyloid-like deposits were associated with the granular deposits usually described in LCDD (Figures 4, 5). To our knowledge there are only four reported cases of such an association: in one case of LCDD reported by Hoffman-Guilaine et al.,5 ultrastructural study showed fibrillar and granular deposits in the walls of renal small vessels. Alpers et al.6 described a polyclonal light chain glomerulopathy with amyloid-like deposits. Recently Kirkpatrick et al.7

Figure 4 Renal biopsy – an electron photomicrograph shows the presence of granular (lighter arrow) and fibrillar (darker arrow) deposits. Magnification ×18,200.

Figure 5 Renal biopsy – closer view of electron-dense granular deposit (left) and of randomly oriented microfibrils (right). Magnification ×56,000.
reported a case of multiple myeloma with kappa light chain deposits in some organs and amyloid deposits in others, the kidney was the only organ to show both amyloid and light chain deposits. Smith and Malcolm,8 however, found in a patient with myeloma, amyloid deposits in hepatic arteries and lambda light chain deposits in the sinusoidal spaces.

In fact, although ultrastructural aspects of LCDD and amyloid deposits are quite different, there are some similarities between LCDD and AL-type amyloidosis: the amyloid-fibril protein is a homogeneous intact immunoglobulin light polypeptide chain whose cellular source is probably an immunocyte-derived clone.9 So, in our patient, light chain and amyloid deposit could be two different expressions of the same immunocyte derived abnormality in protein synthesis. Clinical and biological signs of liver involvement observed in amyloidosis and LCDD were similar.10,11 Using light micro-

References

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