Intestinal electrical activity: a means of assessing viability

E. Q. ARCHAMPONG
B.Sc., M.B., F.R.C.S. (Eng. & Edin.)
Department of Surgery,
Ghana Medical School,
Accra, Ghana

STANISLAV REINIS
M.D., Ph.D.
Department of Physiology,
Ghana Medical School,
Accra, Ghana

Summary
Experiments on ten dogs confirm the impression that electromyographic activity is an important criterion of viability of the intestine subjected to vascular insults.

In all animals gangrene occurred only with complete loss of electrical activity; minimal activity (0.3 mV) was compatible with recovery.

For acceptance in clinical practice it will be essential to prove by further experimental work that an electrically inactive bowel is indeed gangrenous.

Introduction
It is accepted practice to assess the activity and viability of the myocardium, the brain and skeletal muscle, respectively, with the aid of electrocardiography, electroencephalography and electromyography. Extension of this principle to the investigation of the smooth muscle of the intestine has been less enthusiastic although intestinal electromyography has been intensively studied in the past (Alvarez & Mahoney, 1922; Bozler, 1938, 1939, 1942, 1945, 1949; Ambache, 1947; Milton & Smith, 1956; Holaday, Volk & Mandell, 1958; Bass, Code & Lambert, 1961).

To Schamaun (1966) must be attributed the first scientific demonstration of the application of electromyography to assess viability of the bowel. Much, however, remains to be clarified in the practical use of this investigative procedure. The basis of this preliminary report is a study aimed at establishing electromyographic criteria of intestinal viability.

Methods
Electromyographic studies were made on ten dogs weighing between 9 and 30.2 kg. Anaesthesia was with pentobarbitone, 35 mg/kg body weight, and maintained by additional doses of the drug. Three pairs of segments of intestine each 40 cm long in the jejunum, mid-ileum and terminal ileum, respectively, were delimited and their vascular pedicles dissected out, so that the segments were effectively isolated. The artery alone in one of a pair of segments was dissected out and both artery and vein defined in the other.

For recording, fine silver chloride needles (0.4 mm) connected to insulated leads were employed. One needle was inserted into the intestinal wall sub-serosally and secured by means of a stitch; a second needle was fixed to the muscle of the abdominal wall. Recordings were made both on the oscilloscope (Tectronix 502A) and an adapted ECG machine (Cardiovie). In the latter the intestinal pick-up needle was connected to the left leg lead and the abdominal electrode to the left arm lead.

The artery alone was cross-clamped for periods of 4–20 hr and both vein and artery for 4–24 hr. In two dogs intestinal obstruction was produced by tying off a loop of bowel in one segment. Monopolar electromyograms before, and at various stages during and after release of the clamp were obtained. Influences of temperature and pH were studied by bathing the bowel in physiological saline at temperatures between 40° and 20°C and immersion in buffer-solution of pH 3 to pH 9.

Results
The characteristic basic electrical rhythm of Bass et al. (1961) was obtained and found to vary in frequency from 21/min at the upper jejunum to 8/min in the terminal ileum but at any site was constant. This is in agreement with the findings of Milton & Smith (1956).

Effects of vascular occlusion
Application of the clamp to the artery resulted in blanching and shrinkage of the segment, maintained until inspection 4 or 20 hr later. Electromyographic activity showed decrease in frequency and amplitude (Table 1) but never really disappeared (Fig. 1). After release of the clamps all segments recovered their colour and electrical activity was almost completely restored.

Oclusion of artery and vein produced rapid engorgement and darkening of both bowel and mesentery. There was some variation between the
TABLE 1. Effect on electromyographic activity of cross-clamping the artery in segments of dog intestine

<table>
<thead>
<tr>
<th>Segment</th>
<th>Period of occlusion (hr)</th>
<th>Frequency of activity before clamping (per min)</th>
<th>Frequency of activity after clamping (per min)</th>
<th>Frequency of activity after release of clamp (per min)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Mean 17-9 2-6 mV</td>
<td>Mean 14-1 2-3 mV</td>
<td>Mean 14-0 2-3 mV</td>
</tr>
<tr>
<td>1</td>
<td>4</td>
<td>17-4–18-5(20)</td>
<td>13-8–14-4(25)</td>
<td>13-6–14-4(10)</td>
</tr>
<tr>
<td>2</td>
<td>12</td>
<td>17-6–18-4(20)</td>
<td>11-8–12-6(15)</td>
<td>11-0–12-5(15)</td>
</tr>
<tr>
<td>3</td>
<td>20</td>
<td>17-4–18-6(15)</td>
<td>8-5–9-4(10)</td>
<td>6-5–8-8(10)</td>
</tr>
</tbody>
</table>

Fig. 1. Effect of cross-clamping of artery alone in segments of dog ileum 30 cm from duodeno-jejunal flexure. A, Before clamping; B, after clamping for 4 hr; C, after clamping for 12 hr; D, 30 min after release of the clamp.

Fig. 2. Effect of cross-clamping of artery and vein in segments of dog ileum 30 cm from duodeno-jejunal flexure. A–D as in Fig. 1.

TABLE 2. Effect on electromyographic activity of cross-clamping both artery and vein in segments of dog intestine

<table>
<thead>
<tr>
<th>Segment</th>
<th>Period of occlusion (hr)</th>
<th>Frequency of activity before clamping (per min)</th>
<th>Frequency of activity after clamping (per min)</th>
<th>Activity 1 hr after release of clamp (per min)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Mean 18-1 2-4 mV</td>
<td>Mean 11-2 0-6 mV</td>
<td>Mean 10-6 1-0 mV</td>
</tr>
<tr>
<td>1</td>
<td>4</td>
<td>17-6–18-6(20)</td>
<td>11-0–11-4(15)</td>
<td>10-2–11-0(15)</td>
</tr>
<tr>
<td>2</td>
<td>12</td>
<td>18-0–18-2(15)</td>
<td>–</td>
<td>7-5–8-0(20)</td>
</tr>
<tr>
<td>3</td>
<td>24</td>
<td>17-8–18-0(15)</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Intestinal obstruction</td>
<td>50</td>
<td>16-0–16-2(15)</td>
<td>4-0–4-5(10)</td>
<td>10-2–10-5(20)</td>
</tr>
<tr>
<td>Intestinal obstruction</td>
<td>48</td>
<td>16-2–16-6(10)</td>
<td>–</td>
<td>–</td>
</tr>
</tbody>
</table>
Intestinal electrical activity

Table 3. Effect of temperature on electromyographic activity

<table>
<thead>
<tr>
<th>Temperature (°C)</th>
<th>Upper ileum Activity of upper ileum (per min)</th>
<th>Lower ileum Activity of lower ileum (per min)</th>
</tr>
</thead>
<tbody>
<tr>
<td>40°</td>
<td>16:0-15:5(10) Mean 15:7 Mean 2:8 mV</td>
<td>12:5-13:0(12) Mean 12:7 Mean 2:0 mV</td>
</tr>
<tr>
<td>33°</td>
<td>12:0-13:5(15) Mean 12:7 Mean 2:0 mV</td>
<td>8:4-9:5(10) Mean 8:9 Mean 1:8 mV</td>
</tr>
<tr>
<td>20°</td>
<td>9:0-10:2(15) Mean 9:6 Mean 1:5 mV</td>
<td>6:0-7:5(15) Mean 6:7 Mean 1:5 mV</td>
</tr>
</tbody>
</table>

Table 4. Effect of pH on electromyographic activity

<table>
<thead>
<tr>
<th>pH</th>
<th>Activity of upper ileum (per min)</th>
<th>Activity of lower ileum (per min)</th>
<th>Guinea-pig: activity of ileum (per min)</th>
</tr>
</thead>
<tbody>
<tr>
<td>7</td>
<td>19:5-20(15) Mean 19:7 3:0 mV</td>
<td>8:5-9:8(15) Mean 9:1 2:0 mV</td>
<td>8:0-8:6(20) Mean 8:3 1:2 mV</td>
</tr>
<tr>
<td>5</td>
<td>11:5-12:5(20) Mean 12:0 1:5 mV</td>
<td>8:0-9:0(12) Mean 8:5 1:0 mV</td>
<td>7:0-7:5(15) Mean 7:3 0:25 mV</td>
</tr>
<tr>
<td>3</td>
<td>8:0-9:5(15) Mean 8:7 1 mV</td>
<td>6:1-7:3(15) Mean 6:7 0:5 mV</td>
<td>6:0-6:5(15) Mean 6:3 0:25 mV</td>
</tr>
<tr>
<td>9</td>
<td>20:0-20:5(15) Mean 20:2 1 mV</td>
<td>8:6-9:4(12) Mean 9:0 1:0 mV</td>
<td>8:2-8:4(15) Mean 8:3 1:0 mV</td>
</tr>
</tbody>
</table>

segments in the speed of deterioration of electromyographic activity. In one dog low-grade activity was demonstrable after 24 hr of occlusion and the bowel recovered on release of the clamp but in all the other segments occlusion for 12 hr or more produced irreversible loss of electrical activity with gangrene (Fig. 2 and Table 2). In the two dogs with induced intestinal obstruction, the bowel retained a low grade activity (0:3 mV) with complete recovery on relief of obstruction; the other developed gangrene and died.

The effects of temperature and pH are depicted in Tables 3 and 4. Raising the temperature also quickened return of electrical activity after vascular occlusion.

Discussion

The occlusion experiments indicate that small bowel electromyographic activity is particularly sensitive to anoxia. This is not surprising since electrical changes across the cell membrane reflecting ionic exchanges are dependent on an active force provided by cellular metabolism. This confirms Schamaun's (1967) findings.

It is evident further that reduction of activity to very low amplitude (0:3 mV) is compatible with complete recovery. No reproducible patterns of altered activity were noted. In the animals studied infarction was noticed only in complete loss of electromyographic activity. Temperature or pH made no difference to the activity in these cases.

Work is in progress to establish precise quantitative criteria of viability by correlation with histology as well as animal survival. The procedure would find ready clinical application in the investigation of strangulated bowel, mesenteric thrombosis and embolism and trauma. However, for acceptance in clinical practice it would be essential to prove that a section of bowel found to show electrical activity below the accepted criteria is gangrenous. This calls for controlled experiments involving resection and preservation of bowel of critical electrical activity.

References


Intestinal electrical activity: a means of assessing viability

E. Q. Archampong and Stanislav Reinis

*Postgrad Med J* 1969 45: 655-658
doi: 10.1136/pgmj.45.528.655

Updated information and services can be found at:
[http://pmj.bmj.com/content/45/528/655](http://pmj.bmj.com/content/45/528/655)

**Email alerting service**

Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

**Notes**

To request permissions go to:
[http://group.bmj.com/group/rights-licensing/permissions](http://group.bmj.com/group/rights-licensing/permissions)

To order reprints go to:
[http://journals.bmj.com/cgi/reprintform](http://journals.bmj.com/cgi/reprintform)

To subscribe to BMJ go to:
[http://group.bmj.com/subscribe/](http://group.bmj.com/subscribe/)