THE ACTION OF STEROID HORMONES AT THE CELLULAR LEVEL

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Although the complex series of events which occur when a steroid hormone is administered to a human subject or to an intact animal might reflect a series of completely unrelated activities, they are more likely to be secondary to a few fundamental actions on the cells of the body and it is with this second concept that research in this field has been directed.

There are two particular features of steroids which may be important in determining their mode of action. Firstly, they have oxygen substituents at certain typical positions which readily undergo enzymatic oxidoreduction and could, therefore, act as coenzymes or prosthetic groups of enzymes in reactions involving hydrogen transfer. Secondly, steroids are surface active and interact with hydrophobic surfaces producing energy. If the energy were to be absorbed by a receptor molecule it could modify the structure and hence the biological activity of that molecule. A less well defined change is associated with the ability of steroids to capture electrons; this has recently been measured by Lovelock, Simmonds and Vanden- heuvel (1963) who consider that the high electron affinities of adrenocortical hormones, a property unusual among organic compounds, might indicate their ability to participate in or control biological oxidative processes.

Laidler and Krupka (1961) compared the association between the steroid and a receptor with the formation of an activated enzyme-substrate Michaelis complex. Entropy and volume changes during activation of enzymes indicate that structural changes occur in the enzyme molecule and such changes might explain the disturbances of membrane permeability in nerve cells associated with structural changes in acetylcholinesterase. A similar mechanism might account for changes induced by steroids in the permeability of cell structures.

In this process parts of the receptor molecules having specific binding properties might be masked, unmasked, created or destroyed; as well as causing a redistribution of bound substrates, an alteration in enzymic properties might result. Kimberg and Yielding (1962) studied the inhibition of pyruvate kinase by oestrogens and related compounds and showed by viscometry and electrophoresis structural changes in the enzyme, without a change in molecular weight. Yielding and Tomkins (1962) have reported a steroid-hormone induced loss of activity of crystalline glutamic dehydrogenase due to disaggregation into subunits functioning as alanine dehydrogenase with an uncovering of pyridine-nucleotide binding sites. These authors have been more concerned to show the possibility of such changes rather than to attach great physiological significance to them. Chemical changes in receptor molecules after association could account for the highly theoretical possibility of the formation of active enzymes from inactive precursors.

That the receptor itself could be an enzyme cofactor has been considered by Scott and Engel (1961) who obtained physical evidence for the interaction between steroid hormones and purine dinucleotides. This interaction between hormone and a coenzyme associated with a postulated change in the structure of the coenzyme might abolish its hydrogen-carrying function, but there was no experimental evidence for this.

Apart from oxidoreductive changes, metabolism of the steroid molecule is not thought to be of physiological importance except as a mechanism for steroid inactivation. However, competition for active sites on enzymes as opposed to association with them could be of importance in influencing steroid metabolism itself.

The various theories which have been proposed to explain the action of steroid hormones at the cellular level will be considered under the following headings:

1. Effects on membrane permeability and active transport.
2. Effects on hydrogen transfer.
3. Effects on enzyme induction and protein synthesis.

Effects on Membrane Permeability and on Active Transport.

Roberts and Szego (1953) found that oestrogens increased the glucose uptake of rat uteri,
and suggested that their primary action might be to facilitate the entry of glucose into the uterine cells. Noall, Riggs, Walker and Christiansen (1957) showed an increased uptake of \( \alpha \)-amino-\( \text{iso} \)-butyric acid (AIB) in immature rat uteri 20 hours after administering \( \text{estradiol} \) and in rat liver 2 hours after giving hydrocortisone. Since this synthetic amino acid is not metabolised, any change in uptake must be due to a change in transport. Halkerston, Eichorn, Feinstein, Scully and Hechter (1960) examined the effects of \( \text{estradiol} \) on the uteri of castrate rats using intravenous \([^{14}\text{C}]\) labelled AIB as well as \([^{14}\text{C}]\) D-xylose; they found no change 1½ hours after estrogen injection, although the metabolic effects on glucose uptake and incorporation of amino acids into protein are detectable after one hour. After six hours there was an increase in the water content of the tissue and a small increase in AIB and xylose accumulation. They concluded that there is no primary change in sugar or amino acid transport and interpreted previous results as secondary effects operating at a later time. On the other hand Noall and Allen (1961) obtained more than 100 per cent increase in AIB uptake in uteri \textit{in vitro} removed only 30 minutes after intravenous administration of \( \text{estradiol} \) to immature rabbits. They comment that \textit{in vivo} experiments were unsuitable for the investigation of early changes. No effect was observed when the excised uteri were treated with \( \text{estradiol} \) in spite of adequate penetration of the hormone. Although this suggested that the increased uptake observed after \textit{in vivo} hormone administration may not have been a primary effect, they postulated that \( \text{estradiol} \) might be metabolised to a hypothetical active form but their evidence was inconclusive. An increased uptake of AIB by rat levator ani muscles after large doses of testosterone and of synthetic anabolic steroids was shown by Metcalf and Gross (1960) but the effects were not observed until 39 hours after the injection of the steroid and nine hours after AIB administration. Increased AIB uptake in isolated perfused rat livers was seen by Bass, Chambers and Richtarick (1963) two hours after both \textit{in vivo} and \textit{in vitro} administrations of hydrocortisone, suggesting an early direct effect on liver cells.

It is reasonable to conclude that steroid hormones alter transport mechanisms and thus influence the availability of substrate to cells, but although early effects have been observed in isolated organs it is not certain that these are primary events. Since cells are now regarded not as bags of assorted enzymes but as highly organised structures, parts of which are clearly differentiated by membranes and phase boundaries, a possible effect on intracellular substrate distribution must also be considered. Information on this, however, is scanty and indirect. Binding to subcellular particles has been studied by Westphal (1961) who demonstrated interaction between liver mitochondria and hydrocortisone and corticosterone, and by Bellamy (1963) who showed a significant binding between corticosterone and rat liver ribosomes. There is definite evidence that the mitochondrial membrane is influenced by steroids. Westphal (1961) found that hydrocortisone and corticosterone increased the swelling of rat liver mitochondria, indicating an increased permeability to water. Gallagher (1960) concluded that hydrocortisone inhibited oxidative metabolism in liver mitochondria by increasing the membrane permeability causing loss of respiratory cofactors. Blecher (1962) suggested that a similar swelling was associated with a release of latent ATPase which inhibited metabolism of glucose by reducing the availability of ATP. He cited this as a mechanism for the lymphocytolytic action of steroids.

Structural alterations in the peripheral cell membrane are generally assumed to account for changes in transport (see Tomkins and Maxwell, 1963). Controlled and variable passive diffusion can be explained on a biochemical basis, but explanations of mechanisms of active transport in which energy is used to transfer molecules against concentration gradients are only speculative. Hechter and Lester (1960) present and review data suggesting that part of the increased glucose uptake of muscle cells in response to insulin is the apparent removal of intracellular barriers to diffusion, enabling the sugar to equilibrate in a larger fraction of the cell water. Extending this theory the authors outline a model cell based on the results of their experiments on sodium and potassium distribution in the mould \textit{Neurospora Crassa}. Potassium was taken up against a concentration gradient and sodium was excluded from the major part of the cell water, but in the presence of desoxy corticosterone both ions were distributed in all of the cell water at the same concentration as in the surrounding medium. This suggested that the sodium pump operated not at the cell boundary but in the cytoplasm. The crystal structure of certain silicate minerals excludes the highly hydrated sodium ions, whereas the less hydrated potassium ions are able to diffuse freely through and be adsorbed by the lattice.
such minerals behaved in a similar manner to untreated Neurospora. The authors assumed that in the cytoplasm there are ordered lattices which require energy to maintain their structures and which bind potassium ions and water molecules and exclude sodium ions. In the absence of energy or under the influence of specific agents the lattices expand, potassium binding sites are lost and sodium is no longer excluded. Other cytoplasmic components may also bind sugar and amino acid molecules. The bound substrate equilibrates with substrate in the non-bound intracellular water and at equilibrium the concentration of this equals that of the surrounding medium. Active transport in this model is the binding of substrate and water to the lattice with equilibration of the unbound intracellular phase with the surrounding medium through an inert membrane. In spite of the authors’ assertions to the contrary, sufficient binding of potassium to account for active transport is thermodynamically inconsistent with the maintainance of an electrical potential across the membrane. However, the ability of this ingenious model to account for many aspects of active transport merits serious consideration.

Effects on Hydrogen Transfer

Several steroids inhibit the transfer of hydrogen from the pyridine nucleotide coenzymes by the mitochondrial cytochrome chain (see Wade and Jones, 1956) reducing respiration and oxidative phosphorylation. Yielding, Tomkins, Munday and Cowley, (1960) located the site of action between flavoprotein and cytochrome b; Chance and Hollunger (1963) found that inhibition of respiration by progestosterone was sensitive to reagents such as dinitrophenol which uncouple the oxidative and phosphorylation processes, and concluded that the actual site of inhibition was in the energy-transfer process. They postulated that the inhibitor binds a high-energy intermediate and that the uncoupling reagent dissipates this complex with loss of its energy.

Oestrogens stimulate respiration in their target tissues. Hagerman and Villee (1957) studied placental respiration and identified an oestrogen-sensitive isocitric dehydrogenase. This Krebs cycle enzyme requires NADP as coenzyme, but it was at first thought that the oestrogen-sensitive enzyme utilised NAD; later the oestrogen-sensitive step was discovered to be the transfer of hydrogen from NADPH to NAD (Talalay, Hurlock and Williams-Ashman 1958), the rate-limiting factor in the Krebs cycle being the availability of NADP: —

Transhydrogenases, which catalyse the transfer of hydrogen between pyridine nucleotide coenzymes, have been identified in both mitochondria and in soluble cell sap of several tissues and organisms. The transfer of hydrogen from NADPH to NAD increases the efficiency of energy production since NADH transfers its hydrogen to cytochrome c via flavoprotein, coenzyme Q and cytochrome b with a greater yield of ATP than NADPH which transfers hydrogen directly to cytochrome c. They may also play important regulatory roles in cellular metabolism, as NADP may be a rate-limiting factor in dehydrogenations in the pentose phosphate cycle and the availability of NADPH may influence fatty acid and steroid biosynthesis and hydroxylations. In addition to the oestrogen-sensitive transhydrogenase, which Hagerman and Villee (1961) claim can be found in all oestrogen target organs, Hurlock and Talalay (1958) have described a 3-hydroxy-steroid-sensitive transhydrogenase system in liver, but which is probably only of minor importance (Stein and Kaplan, 1959). Pesch, Piros, and Klatskin (1962), and McGuire and Pesch (1962) have identified steroid-sensitive transhydrogenases in particulate preparations of liver and anterior pituitary.

The mechanism of steroid-sensitive transhydrogenation is the subject of some dispute. Talalay and Williams-Ashman (1958) proposed a dehydrogenase system having dual nucleotide specificity with the transfer occurring via the steroid which is alternately reduced and oxidised: —
phoresis they claimed to separate dehydrogenase activity from transhydrogenase activity, although in small yield. Williams-Ashman, Jarabak, Adams and Talalay (1962) purified the placental enzyme 2,500-fold with an overall yield of 29 per cent and could not separate dehydrogenase from transhydrogenase activity at any stage. Moreover the stilbestrol derivatives, having only phenolic oxygen groups, should be inactive if the dehydrogenase theory is correct. Adams, Jarabak and Talalay (1962) reported that with the purified enzyme, diethylstilbestrol acted as a competitive inhibitor. However Abe, Hagerman and Villee (1964) working with an enzyme isolated from myometrium found only slight dehydrogenase activity and that diethylstilbestrol was fully active in stimulating transhydrogenation. Whatever the mechanism, estrogen-stimulated transhydrogenation undoubtedly occurs, and Joel, Hagerman and Villee (1961) present data supporting, but not proving, the hypothesis that increased ATP production after stimulation of transhydrogenation directly influenced the synthesis of cell constituents.

Lucas, Neufeld, Utterback, Martin and Stotz (1955) reported that estrogens stimulate uterine peroxidase activity and Williams-Ashman, Cassman and Calvins (1959) have found a possible hydrogen transfer by purely phenolic compounds in model systems utilizing peroxidase. Talalay and Williams-Ashman (1960) have isolated such a system from uteri and shown an estrogen-mediated increase in oxidation, but its physiological significance is doubtful.

Bush (1962) considers that chemical changes in steroid molecules are not related to their function and represent metabolism and inactivation only. He has examined particularly the 11-oxygen function and from a study of hydrocortisone analogues has concluded that the 11β-hydroxyl group is the only essential requirement for glucocorticoid activity; particularly significant is the finding that 9α-fluorohydrocortisone does not give rise to 11-oxo metabolites, and it therefore seems unlikely that oxidation plays any part in its biological activity since this fluoro compound is many times more active than hydrocortisone.

Effects on Enzyme Induction and Protein Synthesis

The administration of estrogens to females causes a rapid increase in uterine weight and administration of androgens to males causes an increase in the weights of the secondary sex organs and of other sensitive tissues. The increase in weight is due largely to protein although in the uterus there is also a rapid uptake of water. Glucocorticoids cause a net decrease in peripheral-tissue protein whilst appearing to enhance protein synthesis in the liver. In addition to the general weight response of target tissues to estrogens or androgens, administration of these hormones increases the effective concentration of many enzymes and glucocorticoids increase the effective concentration of some liver enzymes. The response to androgens and probably to other steroid hormones is highly dependent on both tissue and species.

*Induced enzyme synthesis.* The effect of steroid hormones on protein synthesis is most easily studied by determining changes in enzyme activity because of the ease with which small net changes may be measured. An increase in activity of an enzyme after steroid administration could be due to either enzyme activation or an increase in enzyme synthesis, i.e. enzyme induction. Induction, however, might not necessarily be a primary action of the steroid but could be secondary to some other action of the steroid in modifying the intracellular environment. Much experimentation has been designed to establish whether there is activation or induction, often by immunological studies.

The effect of testosterone on the activity of mouse kidney β-glucuronidase has been studied by Riotton and Fishman (1953). The activity of this enzyme after treatment of the animal with testosterone showed a thirtyfold increase and could be maintained at this level for 60 days by continued testosterone administration. The response was later attributed to an increase in enzyme synthesis because the specific activity of β-glucuronidase isolated from the kidneys of mice treated with testosterone and given [14C]-glycine was higher than that from the kidneys of untreated mice or the livers of both treated and untreated animals (Pettengill and Fishman, 1960). Tryptophan pyrrolase activity in liver is stimulated by the administration of hydrocortisone to rats but this increase is prevented by simultaneous administration of the methionine analogue ethionine (Horton and Franz, 1959); in perfused-liver preparations the increase occurs after the hydrocortisone has disappeared (Goldstein, Stella and Knox, 1962). The increase in enzyme activity is, therefore, a result of increased enzyme synthesis.

Hydrocortisone produces a rapid increase in tyrosine-α-ketoglutarate transaminase activity in perfused rat-liver preparations (Goldstein
and others, 1962). Kenney (1962a) has purified this enzyme and shown that there are both active and inactive enzyme components but the proportions of active and inactive components were the same in preparations from both control and hydrocortisone-treated animals. He concluded that hydrocortisone did not promote the conversion of an inactive enzyme form to the active form and he also showed (Kenney 1962b) that the rise in enzyme activity was paralleled by a rise in antigenic material suggesting that increased enzyme synthesis was primarily responsible for the hydrocortisone-induced increase in activity. Kenney and Flora (1961) showed that the increase in activity was related to the concentration of steroid in the liver and believed that the steroid was acting directly on the protein-synthesising system.

Glutamic-pyruvic transaminase activity in rat liver is increased by hydrocortisone administration and this increase could be inhibited by ethionine (Segal, Beattie and Hopper, 1962). Segal, Rosso, Hopper and Weber (1962) have shown that the rise in enzyme activity is paralleled by an increase in antigenic material again suggesting that the hormone induces enzyme synthesis. However, fasting, diabetes or a high protein diet also increases hepatic glutamic-pyruvic transaminase activity (Rosen, Roberts and Nichol, 1959) which suggests that the observed increase may be secondary to a peripheral effect. Similarly, although hydrocortisone increases glucose-6-phosphatase activity in the livers of adrenalectomized or normal rats so does starvation or a high protein diet (Harper and Young, 1959; Harper, 1959) or alloxan diabetes (Fresch, Ashmore and Renold, 1958).

**Effects on uptake of amino acids.** Several workers have studied the uptake of radioactive amino acids by target tissues after oestrogen or androgen administration. Frieden, Laby, Bates, and Layman (1957) showed that the uptake of [14C]-glycine into mouse kidney was stimulated by testosterone propionate and Butenandt, Gunther and Turba (1960) showed that the incorporation of [14C]-leucine by seminal vesicles of immature rats receiving testosterone was increased fourfold whilst Mueller (1961) has shown that the amount of radioactivity in protein was greater in uteri from oestradiol-treated rats after [14C]-glycine administration. Mueller (1961) also showed that the incorporation of radioactivity from [14C]-glycine into nucleic acid adenine and of 32P into phospholipid of uteri was increased when rats were treated with oestradiol. Puromycin inhibited these effects of oestradiol. He concluded that oestradiol-stimulated protein synthesis and that the effect of puromycin on the incorporation of radioactivity from glycine into nucleic acid adenine and of phosphorus into phospholipid was due to an inhibition of the enzymes responsible for adenine and phospholipid synthesis. Such experiments do not define the step in protein synthesis on which steroids act.

Wilson (1962a) in an attempt to define this more closely has studied the rise in specific activity of free intracellular tyrosine and of protein during incubation of slices of rat seminal vesicles with [14C]-tyrosine. He concluded that testosterone did not act by enhancing amino acid transport and that the increase in amino acid incorporation was secondary to an increased protein synthesis.

**Effects on amino acid activation.** Steroids may not only act directly on amino acid transport (see above) but have been shown to increase also the activity of some amino-acid-activating enzymes, which are essential for the synthesis of protein. Thus, Mueller, Herranen and Jervell (1958) have shown that oestradiol causes an early increase in the activity of seven of the known amino-acid-activating enzymes and Kochakian, Tanaka and Hill (1961) have shown that the activity of amino-acid-activating enzymes in the seminal vesicles of guinea pigs was reduced 40 per cent 40 days after castration. The activity could be restored by androgens.

**Effects on ribosomal activity and the formation of ribonucleoprotein.** Much recent work on steroids has been designed to define their exact role in protein synthesis, the main steps of which are outlined in Figure 1.

Amino acids are activated in the cytoplasm by specific amino-acid-activating enzymes and can then complex with specific soluble ribonucleic acid molecules (s-RNA). The s-RNA-amino-acid complex which is formed is transferred to a specific position on an RNA template situated on the surface of a ribosome. Peptide-bonding of the amino acids attached to the template occurs and is followed by release of the newly formed protein from the template. The genetic determination of the structure of a protein is due to the synthesis of that protein on a ribosomal RNA template whose base sequence is determined by that of the deoxyribonucleic acid (DNA) of the structural gene. The template or messenger RNA is synthesised in the nucleus and transferred to a ribosome; its rate of synthesis is determined by an operator-repressor gene system (Jacob and Monod, 1961). The operator is normally inhibited by the repressor and cytoplasmic inducers act by reducing the inhibitory action of the repressor on the operator. Since the ability of ribosomes to synthesise protein is related to its template RNA.
content protein synthesis will be affected. Steroids could act on protein synthesis by regulating the availability of template RNA through control of its rate of synthesis or breakdown, or alternatively by modifying ribosomal structure as Tissières, Schlesinger and Gros (1960) have shown that this is important in determining the rate of protein synthesis in bacterial systems.

Wilson (1962a) investigated the effect of testosterone administration on the increase with time in specific activity of ribosomal protein and s-RNA-amino-acid complex in slices of seminal vesicle incubated with [14C]-tyrosine or [14C]-leucine. He found that testosterone accelerated the rise in specific activity of ribosomal protein but did not affect the rise in specific activity of the s-RNA-amino-acid. He concluded that testosterone acted by enhancing the formation of ribonucleoprotein on the ribosomes and in similar experiments on the effect of oestradiol on protein synthesis in rat uterine slices, Wilson (1962b) showed that oestradiol also acted on this step. Using preparations of ribosomes and soluble factors isolated from the oviducts of oestradiol-treated or control hens, Wilson (1962b) showed that oestradiol modified ribosomal activity but did not affect the activity of the soluble factors required in the formation of ribonucleoprotein.

The action of glucocorticoids on protein synthesis in the liver has also been defined at the ribosomal level by Korner (1960) who showed that ribosomes from the livers of rats adrenalectomized some weeks previously were less active than those of controls. Treatment of normal rats with hydrocortisone increased liver ribosome activity. The immediate effect of adrenalectomy, however, was to increase ribosomal activity and this could be abolished by treatment with hydrocortisone. The activity of the soluble factors was unchanged during these procedures. These paradoxical early and late effects of adrenalectomy have not yet been explained.

**Effects on synthesis of template (ribosomal) ribonucleic acid.** Mueller (1961) has shown that increased nucleic acid synthesis is an early effect of oestrogen treatment; Kochakian, Hill and Aonuma (1963) have found that template RNA from mouse kidney, but not soluble RNA, was increased by testosterone propionate administration and Cantarow, Pashkiss and Williams (1959) have found that the administration of testosterone increased [14C]-uracil incorporation by rat liver and hepatoma cells. More directly, Hancock, Zelis, Shaw and Williams-Ashman (1962) have shown that the incorporation of radioactive cytosine triphosphate into RNA by DNA-dependent RNA polymerase from the nuclei of rat ventral prostate was decreased in preparations from castrated rats and the activity could be restored by treating the rats with testosterone. These experiments show that steroids increase the synthesis of template RNA.

Work on the inhibition of induced enzyme synthesis by 8-azaguanine (which prevents RNA synthesis) and by actinomycin (which prevents the messenger RNA from attaching to the ribosomes) has shown that the synthesis of template RNA and its attachment to the ribosomes are necessary for induced enzyme synthesis. Thus, Kvam and Parkes (1960) have shown that the simultaneous administration of 8-azaguanine and hydrocortisone to adrenalectomized rats prevented the increase in glucose-6-phosphatase and fructose 1, 6-diphosphatase which normally occurred after hydrocortisone administration. Similarly Greengard and Acs (1962) showed that actinomycin prevented the rise in tryptophan pyrrolase and tyrosine-α-ketoglutarate transaminase activity after hydrocortisone administration. However, chloramphenicol, which inhibits protein synthesis in bacteria and is thought to act by preventing the messenger RNA from attaching to the ribosomes, does not inhibit the incorporation of [14C]-leucine into kidney ribosomes from testosterone-treated mice (Kochakian and others, 1963) nor does it inhibit protein synthesis in the oviduct system of hens treated with oestradiol (Wilson, 1962b). This does not invalidate the
view that steroids act by increasing the synthesis of template RNA, but shows that there is not a high turnover of labile messenger RNA sensitive to chloramphenicol inhibition as in bacteria.

Direct evidence that steroids act on protein synthesis by regulating the availability of template RNA has been obtained by Liao and Williams-Ashman (1962). They found that the transfer of radioactivity from s-RNA-valine-\[\text{\textsuperscript{14}C}\] to trichloracetic acid-insoluble material by ribonucleoprotein particles from rat ventral prostate was markedly reduced in preparations from animals castrated 2-3 days previously and the activity could be increased threefold by the addition of polyuridine-guanidine (UG), a synthetic polynucleotide having a high uracil/guanine ratio. The administration of testosterone to the animals completely reversed the effects of castration and the addition of poly-UG to preparations from testosterone-treated castrates had no effect. On the other hand, Wilson (1962b) found that the addition of ribosomal RNA, extracted from the oviducts of hens treated with oestradiol, to the protein-synthesising system from the oviducts of untreated hens did not increase the rate of protein synthesis. However, the RNA might have been degraded during the extraction procedure.

Effects of glucocorticoids on protein synthesis in peripheral tissues. The glucocorticoid-induced decrease in peripheral-tissue protein seems to be due to a decreased rate of synthesis rather than to an increased rate of breakdown. Manchester, Randle and Young (1959) showed that adrenalectomy raised, whilst hydrocortisone depressed, the rate of incorporation of \[\text{\textsuperscript{14}C}\]-glycine into protein of rat diaphragm and White, Hoberman and Szego (1948) showed that the rate of incorporation of \[\text{\textsuperscript{14}N}\]-glycine into muscle and some other tissues was increased by adrenalectomy. From a study of the rate of excretion of isotopic nitrogen after the administration of \[\text{\textsuperscript{14}N}\]-glycine to normal, adrenalectomized and adrenalectomized rats treated with cortisone, Clarke (1953) concluded that cortisone acted by inhibiting synthesis rather than by promoting protein breakdown.

Conclusions

Many of the physiological effects of steroids on electrolyte distribution and on the availability of cellular substrates might result from a single action of these hormones on active transport. Although the exact mechanism of active transport is unknown, the theory of Hechter and Lester is interesting. Steroids could interact with those intracellular structures which are involved in the transport mechanism, modify their structure and hence the uptake of substances by the cell. However, the uptake of substrate by cells is probably not rate-limiting in all cases and steroids certainly affect the activity of many intracellular enzyme systems so that an apparent action on transport could be secondary to some other action on an intracellular enzyme system.

Steroid-activated enzyme systems which catalyze transhydrogenation occur in many tissues but their presence in all oestrogen target tissues is questionable (Mueller, 1961) which throws doubt on their physiological significance. The mechanism proposed by Talalay for transhydrogenation is the only example of chemical change in the steroid molecule being important in its mode of action.

The action of steroids on protein synthesis, with the possible exception of the influence of glucocorticoids on peripheral tissues, is probably to regulate the synthesis of messenger RNA, perhaps through the operator-repressor gene system. Such a mechanism permits specific enzyme synthesis and a differentiation in tissue response because in different tissues enzymes with similar functions may have different structures (Kaplan, Ciotti, Hamolsky and Bieber, 1960) and hence be controlled by different gene complexes. However, the increase in synthesis of some enzymes after steroid administration might not be due to a direct action of the steroid on the protein-synthesising mechanism but could be secondary to some other change in cellular environment.

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