Neurological channelopathies

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In order for cells to retain their integrity to water and yet permeate charged ions, transmembrane proteins known as ion channels have evolved. There is huge diversity of these ion channels. Some proteins are tissue specific, while others are widely distributed throughout the body. The resting membrane potential of excitable cells is entirely due to the presence of such ion channels. It is therefore unsurprising that these channels are integral to the fundamental processes of electrical signalling and excitation within the nervous system. It had been suspected that genetic dysfunction of such critical membrane-bound proteins would be lethal. However, during the past few years there has been an explosion in the discovery of disease-causing mutations in genes coding for ion channel proteins and these disorders are known as channelopathies. We now recognise both genetic and autoimmune channelopathies affecting a range of tissues. This review considers clinical, genetic, autoimmune, and molecular pathophysiological features of the neurological channelopathies.

CLASSIFICATION OF ION CHANNELS

Different classifications of ion channels exist. For the purpose of this review we have classified ion channels into two broad categories depending on their mode of activation—that is, voltage gated and ligand gated. Table 1 shows how the genetic neurological channelopathies are subdivided on the basis of channel type. Table 2 is a list of genetic neurological channelopathies according to ion type. Most ion channels have a similar basic structure. All voltage gated ion channels have a large pore forming subunit, which sits within the membrane. The pore forming subunit (also called the α-subunit) contains a central aqueous pore through which the relevant ion passes in response to voltage change induced activation, also known as gating. In addition to the main α-subunit, it is common for voltage gated ion channels to possess accessory subunits, these subunits may be cytoplasmic or extracellular. Generally, these have an important function in modulating the basic conductance function of the α-subunits. The structural topology of all voltage gated ion channels is remarkably conserved through evolution. To date, most genetic neurological channelopathies affecting the peripheral nervous system (PNS) and central nervous system (CNS) are caused by α-subunit mutations, resulting in dysfunction of voltage gated ion channels. However, examples of genetic channelopathies due to dysfunction of ligand gated channels are recognised, particularly in the PNS and are emerging in the CNS. To date most autoimmune channelopathies affect the PNS, although CNS examples are likely to increase in the future.

INHERITED CHANNELOPATHIES

Muscle channelopathies

Myotonic syndromes

Myotonia is the term given to delayed relaxation of skeletal muscle after voluntary contraction. In most situations myotonia is most marked after initial muscle contraction, and usually abates after repeated muscle activity (the warm-up phenomenon). Electrophysiologically, myotonia is a disturbance of the normal excitability of the skeletal muscle membrane. There is an abnormally increased excitability of the membrane such that in response to a depolarising stimulus, for example, a nerve impulse, rather than a single muscle contraction being initiated, multiple contractions occur and this results in the delayed relaxation observed clinically.

From a practical point of view a myotonic disorder should be considered in the differential
diagnosis of a patient complaining of muscle stiffness. Myotonic dystrophy is a common and important cause of myotonia but the presence of extramuscular systemic symptoms and signs usually aid the diagnosis. The pure myotonic disorders considered here do not cause multisystem disease. For these disorders, particular attention should be paid to any family history and to the precipitants of the muscle stiffness, for example, temperature and whether the patient's stiffness reduces with exercise—the so called warm-up phenomenon or whether stiffness increases with exercise—so called paradoxical myotonia (see below). Furthermore the presence of muscle hypertrophy should be sought on examination and may be a clue to a chloride channel myotonia.

**Myotonia congenita**

See table 3 for clinical features.

**Thomsen’s disease**

Dr Thomsen initially described this in his own family in 1876. Patients usually present between infancy and adulthood with mild myotonia, which may be constant or intermittent. Marked improvement in myotonia is noted with repeated exercise of a given muscle, the warm-up phenomenon. While 90% show myotonia on electromyography, only 50% have percussion myotonia on examination. There is usually normal power at rest, although some have proximal weakness, which can present with functional difficulties such as climbing stairs. Some patients have muscle hypertrophy while others complain of myalgia. Electromyography shows myotonia with a distal predominance, which is present even in early childhood and the warm-up effect can be observed electrophysiologically.

Thomsen’s disease is caused by mutations in a muscle voltage gated chloride channel (CLCN1) located on chromosome 7q35. It is transmitted as an autosomal dominant trait with variable penetrance, although 90% of affected individuals are symptomatic. This channel exists as a dimer, mutations may interfere with dimerisation by exerting a dominant negative effect on the wild-type subunits. Since chloride conductance is necessary to stabilise the high resting membrane potential of skeletal muscle, the loss of chloride conductance caused by mutations results in partial depolarisation of the membrane allowing increased excitability and myotonia.

### Table 1

<table>
<thead>
<tr>
<th>Muscle</th>
<th>Central nervous system</th>
</tr>
</thead>
<tbody>
<tr>
<td>Voltage gated channels</td>
<td></td>
</tr>
<tr>
<td>Hypokalaemic periodic paralysis</td>
<td>Episodic ataxia type 1</td>
</tr>
<tr>
<td>Hyperkalaemic periodic paralysis</td>
<td>Episodic ataxia type 2</td>
</tr>
<tr>
<td>Andersen’s syndrome</td>
<td>Familial hemiplegic migraine</td>
</tr>
<tr>
<td>Myotonia congenita</td>
<td>Several inherited epilepsy syndromes</td>
</tr>
<tr>
<td>Paramyotonia congenita</td>
<td>Malignant hyperthermia</td>
</tr>
<tr>
<td>Chloride channel</td>
<td>Hypokalaemic periodic paralysis</td>
</tr>
<tr>
<td>Myotonia congenita: Thomsen’s, Becker’s</td>
<td>Episodic ataxia type 2</td>
</tr>
<tr>
<td>Calcium channel</td>
<td>Hypokalaemic periodic paralysis</td>
</tr>
<tr>
<td>Malignant hyperthermia</td>
<td>Familial hemiplegic migraine</td>
</tr>
<tr>
<td>Potassium aggravated myotonia</td>
<td>Childhood absence epilepsy</td>
</tr>
<tr>
<td>Potassium channel</td>
<td>Andersen’s syndrome</td>
</tr>
<tr>
<td>Hypokalaemic periodic paralysis</td>
<td>Episodic ataxia type 1</td>
</tr>
<tr>
<td>Hyperkalaemic periodic paralysis</td>
<td>Benign familial neonatal convulsions</td>
</tr>
<tr>
<td>Ryanodine receptor</td>
<td>Malignant hyperthermia</td>
</tr>
<tr>
<td>Central core disease</td>
<td>Epilepsy</td>
</tr>
<tr>
<td>Glycine receptor</td>
<td>Hyperkplexia</td>
</tr>
<tr>
<td>Acetylcholine receptor</td>
<td>Autosomal dominant focal lobe epilepsy</td>
</tr>
<tr>
<td>GABA receptor</td>
<td>GEF5F, juvenile myoclonic epilepsy</td>
</tr>
</tbody>
</table>

### Table 2

<table>
<thead>
<tr>
<th>Channel</th>
<th>Muscle</th>
<th>Gene</th>
<th>CNS</th>
<th>Gene</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sodium channel</td>
<td>Hypokalaemic periodic paralysis</td>
<td>SCN4A</td>
<td>Generalised epilepsy with febrile seizures plus syndrome (GEFS+), severe myoclonic epilepsy of infancy</td>
<td>SCN1A SCN1B SCN2A</td>
</tr>
<tr>
<td>Calcium channel</td>
<td>Hypokalaemic periodic paralysis</td>
<td>CACNA1S</td>
<td>Episodic ataxia type 2</td>
<td>CACNA1A</td>
</tr>
<tr>
<td>Potassium channel</td>
<td>Andersen’s syndrome</td>
<td>KCN12</td>
<td>Episodic ataxia type 1</td>
<td>KCNA1 KCNQ2 KCNQ3</td>
</tr>
<tr>
<td>Ryanodine receptor</td>
<td>Malignant hyperthermia</td>
<td>RYR1</td>
<td></td>
<td>RYR1</td>
</tr>
<tr>
<td>Glycine receptor</td>
<td></td>
<td></td>
<td>Hyperkplexia</td>
<td>GLRA1</td>
</tr>
<tr>
<td>Acetylcholine receptor</td>
<td></td>
<td></td>
<td>Autosomal dominant focal lobe epilepsy</td>
<td>CHRNA4</td>
</tr>
<tr>
<td>GABA receptor</td>
<td></td>
<td></td>
<td>GEF5F, juvenile myoclonic epilepsy</td>
<td>GABRG2</td>
</tr>
</tbody>
</table>
Becker’s disease

The Becker form of myotonia congenita is more severe than Thomsen’s disease with an earlier age of onset. As in Thomsen’s disease there is myotonia with the warm-up phenomenon but patients also have significant muscle hypertrophy, especially in the gluteal muscles. There may also be mild distal muscle weakness. Strength is normal initially but there may be rapid decrease in power with short amounts of exercise, which returns to normal after further muscle contraction. Such transient weakness in Becker patients is more likely to happen after a period of rest. For example after sitting for a while a patient may experience a transient lower limb weakness on standing. The electromyogram shows frequent myotonic discharges and the warm-up effect can be demonstrated. In contrast to Thomsen’s disease the motor units are frequently mildly myopathic. Becker’s disease is also due to mutations in the muscle chloride channel (CLCN1), hence the two forms of myotonia congenita are allelic. However, Becker’s disease shows autosomal recessive inheritance. There is a male predominance, suggesting reduced penetrance or a milder clinical phenotype in females. Mutations have been found throughout the gene, with missense and nonsense mutations and deletions identified. Most patients are compound heterozygotes. Expression studies have indicated that the majority of mutations result in a loss of function of the chloride channel monomer.²

Practical management

Many patients with myotonia congenita do not require medication, but in our experience those that do usually respond well to mexiletine. Other antymytogenic agents can be considered and include phenytoin, but these are less effective.⁴ Mexilitene causes use-dependent blockade of sodium channels and stops the production of repetitive runs of action potentials and hence reduces muscle stiffness. However, it can lead to arrhythmias, including torsades de pointes and as it is unlicensed in the UK for myotonia and approval should therefore be sought from local use of medicines committees. We suggest that it is only prescribed by neurologists with experience of its use in this context. Mexilitine treatment requires close monitoring with electrocardiography. Ultimately a specific chloride channel opening agent would be the ideal therapy for such patients but such a drug has not been developed to date. Accurate genetic counselling is important, especially with regards to risks to offspring and this relies on the availability of a precise DNA based diagnosis.

Potassium aggravated myotonias

This is an umbrella term for several conditions due to mutations in the skeletal muscle voltage gated sodium channel, SCN4A (described in detail below). Clinically patients exhibit pure myotonia of variable severity, which can be particularly sensitive to potassium ingestion with no associated weakness. Clinically, distinction from myotonia congenita, described above, may be difficult. Various terms have been used to describe these disorders, which are summarised below.

Potassium aggravated myotonia

This is characterised by mild myotonia that varies in severity from day to day with no weakness or cold sensitivity. Stiffness typically develops during rest after a period of exercise and lasts for approximately one hour. It is exacerbated by potassium and depolarising agents (for example, suxamethonium) and may interfere with respiration. The electromyogram shows myotonia which increases after exercise.¹

Myotonia fluctuans

This is characterised by mild myotonia that varies in severity from day to day with no weakness or cold sensitivity. Stiffness typically develops during rest after a period of exercise and lasts for approximately one hour. It is exacerbated by potassium and depolarising agents (for example, suxamethonium) and may interfere with respiration. The electromyogram shows myotonia which increases after exercise.¹

Myotonia permanens

In this condition patients experience severe continuous myotonia, which may interfere with respiration. There is often marked muscle hypertrophy, especially in the neck and shoulders.

Acetazolamide responsive myotonia congenita

This is characterised by muscle hypertrophy, myotonia and myalgia, is aggravated by potassium loading and improved by acetazolamide.⁷

Paramyotonia congenita

“Paradoxical” myotonia is stiffness (myotonia) that appears during exercise and worsens with continued activity. Electromyography at rest often shows some myotonia, although it is often less prominent than in the other myotonias described above. Low temperature often precipitates symptoms in these

| Table 3  Clinical features of myotonia and paramyotonia congenita |
|---------------------------------|-----------------|-----------------|-----------------|
| **Inheritance** | Paramyotonia congenita | Myotonia congenita | Thomsen’s |
| Autosomal dominant | | Autosomal dominant | Autosomal recessive |
| **Age of onset** | Neonatal to infancy | Early childhood | First decade |
| **Anatomical distribution** | Face, tongue, neck, arms | Face, arms, >legs | Legs, >arms, face |
| **Exacerbating factors** | Cold, exertion, spontaneous | Cold, rest, hunger, fatigue, stress | |
| **Clinical features** | Cold induced weakness | Cold induced weakness | Myotonia generally more disabling than in Thomsen’s disease. |
| | usually lasts a few minutes, but occasionally | days some associated | |
| | days some associated | with HyperKPP | |
| **Muscle hypertrophy** | Absent | Present | Present |
| **Cold immersion** | Cold, rest, hunger, fatigue, stress | |
| **Electromyography findings** | CMAP amplitude | No decrement | |
| | CMAP amplitude | | |
| | decrement with cooling | | |
| **Treatment** | Mexilitine,¹ acetazolamide | Mexilitine,¹ acetazolamide,² | phenytoin³ |
| | | | |
| **Ion channel gene** | Sodium channel (SCN4A)¹ | Chloride channel (CLCN1)¹ | |

CMAP, compound muscle action potential; HyperKPP, hyperkalaemic periodic paralysis.
patients and cooling produces repetitive spontaneous motor unit discharges with a decrement in the compound muscle action potential (CMAP) amplitude. The clinical features are summarised in table 3.

Paramyotonia congenita is caused by mutations in the voltage gated skeletal muscle sodium channel α-subunit (SCN4A) on chromosome 17q35. Voltage dependent activation of this channel results in influx of sodium into the muscle fibre and is therefore responsible for the upstroke of the action potential. Rapid closure of this channel after activation is critical for muscle fibre repolarisation. Paramyotonia congenita is inherited as a highly penetrant autosomal dominant trait. Mutations have been found throughout the gene, although exon 24 appears to be a hotspot for mutations. Mild depolarisation (>5 mV) produces repetitive discharges (myotonia) while more severe depolarisation (>20 mV) produces weakness, either of which may occur as an isolated phenomenon.

Treatment
The myotonia usually responds to antiarrhythmic drugs such as mexilitine. The weakness is potassium sensitive and responds to hydrochlorothiazide, acetazolamide or dichlorphenamide, with or without potassium supplementation.

Differential diagnosis of myotonia
See table 4 for the differential diagnosis of myotonia.

Many rheumatological conditions may associate with the symptom of muscle stiffness. There are usually associated clinical clues that point to this correct diagnosis as well joint pains. In our experience some cases diagnosed with chronic fatigue or fibromyalgia have turned out to have myotonic disorders such as myotonia congenita, emphasising the need for careful clinical and electromyography assessment in such cases. A differential diagnosis of neurological conditions resulting in muscle stiffness is given in table 4.

Periodic paralyses
Hyperkalaemic periodic paralysis
Hyperkalaemic periodic paralysis (HyperKPP) is an autosomal dominant disorder with an estimated prevalence of 1:200 000. Patients experience attacks of either focal or generalised muscle weakness often after exercise. Attacks may vary in severity from mild weakness to total paralysis. The duration of attacks is shorter than in hypokalaemic periodic paralysis (HypoKPP) and typically lasts about an hour or two. The attack frequency declines with age but patients often develop a fixed myopathy of variable severity. The clinical features are shown in table 5. It is notable that death is fortunately extremely rare in HyperKPP or HypoKPP. In contrast to Andersen’s syndrome (see below), cardiac arrhythmias are uncommon, as the ion channels mutated in HyperPP and HypoKPP are not expressed in cardiac muscle. HyperKPP is caused by point mutations in the skeletal muscle sodium channel α-subunit, SCN4A (which is mutated in paramyotonia congenita). These mutations lead to defective inactivation of the channel. Some genotype/phenotype correlations can be made. For example, the most frequent point mutation, T704M, which occurs in 60% of cases frequently leads to permanent late onset muscle weakness. Another frequent mutation, I1592M, is often associated with myotonia in addition to paralysis.

Attacks of weakness are associated with high serum potassium and high urinary potassium excretion. However, it is important to note that the serum potassium may remain within the normal range and that hyperkalaemia may rapidly autocorrect, therefore measurement as early as possible during an attack is critical. The creatine kinase may be normal or modestly increased to about 300 U/l. Many attacks are brief and do not require treatment. If necessary, acute attacks can be terminated by ingestion of carbohydrate or inhaled salbutamol. Preventive treatment with acetazolamide or a thiazide diuretic may be required. HyperKPP caused by a mutation in the potassium channel, KCNE3, has been reported only in one family.

Andersen’s syndrome
Andersen’s syndrome is an autosomal dominant potassium sensitive periodic paralysis with ventricular dysrhythmias and dysmorphic features. The dysmorphic features are often subtle but include low set ears, hypertelorism, clinodactyly, and syndactyly. Bidirectional ventricular tachycardia is a frequent and potentially serious arrhythmia. From a practical point of view, this disorder should be considered in any case of periodic paralysis with arrhythmia. The resting electrocardiogram often shows bigeminy. The clinical features are

<table>
<thead>
<tr>
<th>Table 4 Differential diagnosis of neurological conditions that may mimic myotonia</th>
</tr>
</thead>
<tbody>
<tr>
<td>Region affected</td>
</tr>
<tr>
<td>Central nervous system</td>
</tr>
<tr>
<td>Peripheral nervous system</td>
</tr>
<tr>
<td>Muscle</td>
</tr>
<tr>
<td>Metabolic muscle disease</td>
</tr>
<tr>
<td>Ripping muscle disease</td>
</tr>
</tbody>
</table>

EMG, electromyography; PMC, paramyotonia congenita.
summarised in table 6. It is now known that Andersen’s syndrome is a cardioskeletal muscle channelopathy caused by mutations in a potassium channel termed Kir2.1. This inward rectifying potassium channel is encoded by KCNJ2 on chromosome 17q23 and disease-causing mutations were first described in 2001.16 The channel plays a part in cardiac and skeletal muscle membrane hyperpolarisation and interestingly, also has a role in skeletal bone precursor cell migration and fusion during development, hence the triad of symptoms. Functional expression studies have shown loss of function due to a dominant negative effect on wild-type channel subunits, producing a reduced inwardly rectifying K+ current.17 There is intrafamilial variability and partial manifestation of the phenotype is common. Serum potassium during an attack may be high, low, or normal. In those with hypokalaemia, oral potassium supplements may improve the weakness. In some families increasing plasma potassium concentration with acetazolamide improves arrhythmias at the expense of exacerbating weakness.18 Once the diagnosis is made detailed cardiac assessment is essential. However, the optimum management to prevent malignant arrhythmias is not certain. Currently, opinions vary from imipramine treatment to implantable cardioverter defibrillators.

### Hypokalaemic periodic paralysis

HypoKPP is the most common form of periodic paralysis with an incidence estimated to be one in 100 000. It is inherited in an autosomal dominant manner but new mutations account for up to one third of cases. Comparison with HyperKPP is made in table 5. The attacks may be brought on by a period of exercise followed by rest or by carbohydrate loading. It is common for attacks to develop in the early hours of the morning, particularly if a large carbohydrate meal was taken late the previous evening. Serum potassium is typically low at the onset but may normalise quickly. However, there is no correlation between serum potassium concentration and the severity of weakness. The creatine kinase is increased during attacks. Conduction velocity in muscle fibres is slow; CMAPs are reduced during attacks and increase immediately after sustained (five minutes) maximal contraction. As in all forms of periodic paralysis attack frequency tends to decline with age but a fixed myopathy may develop. Myotonia never occurs in HypoKPP.

Point mutations in three separate muscle channel genes may cause HypoKPP. The majority of cases harbour one of three point mutations in the L-type calcium channel, CACNA1S. Far less frequent mutations have been described.
in the muscle sodium channel SCN4A, and in the potassium channel KCNE3. Mutations in the L-type calcium channel α1-subunit (dihydropyridine receptor) (CACNA1S), located on chromosome 1q31, account for about 70% of cases of HypoKPP.13 Mutations in KCNE3 on chromosome 11q13–q14 have only been reported in one family.14 Mutations in the ryanodine receptor (RYR1) on chromosome 19q1325 are found in 50% of families with HypoKPP. All mutations are arginine substitutions in the voltage sensor of the channel protein. It remains unclear how mutations in CACNA1S, which does not have a major role in determining muscle membrane excitability, result in attacks of paralysis. The normal channel has two roles: (1) as a slow voltage activated calcium channel and (2) excitation contraction coupling with the ryanodine receptor. Mutated channels have enhanced inactivation leading to a very small defect in the control of muscle resting membrane potential. There is reduced penetrance in females (50%) compared with complete penetrance in males. About half of the women who have the R528H mutation and one third of those with the R1239H mutation are asymptomatic. In contrast, more than 90% of males with a disease-causing mutation are symptomatic. Specific mutations appear to have discrete clinical features—for example, R528H is common, with later onset and associated myalgias. The other major group of HypoKPP are due to missense mutations in the voltage sensor of domain 2 of SCN4A. (the sodium channel affected in HyperKPP and paramyotonia congenita). There is some genotype/phenotype correlation—for example, acetazolamide treatment is often deleterious in the R672G mutation. SCN4A mutations are an uncommon cause of HypoKPP in the UK. Mutations in KCNE3 on chromosome 11q13–q14 have only been reported in one family.14

### Practical approach to suspected periodic paralysis

A high index of suspicion, an accurate history, neurological examination during an attack, and measurement of serum potassium in a sample taken as early as possible after presentation are the keys to making the diagnosis of periodic paralysis. HypoKPP can also occur in the context of hyperthyroidism (usually in Asian patients), so thyroid function tests should also be measured. Other general medical causes of altered potassium concentrations should always be sought. Associated features which support the suspicion of genetic periodic paralysis may include dysmorphic features (Andersen’s syndrome) and myotonia or paramyotonia. Generally we find provocative tests, such as potassium loading or induction of hypokalaemia, unhelpful and they are potentially hazardous. DNA testing should be considered at an early stage with the patient’s consent (see end of article for details). If diagnostic uncertainty remains referral to a specialist centre should be considered.

### Malignant hyperthermia syndromes

Malignant hyperthermia syndrome is the most common cause of death during anaesthesia, with an estimated incidence of somewhere between one in 7000 to one in 50 000 of anaesthetics given. There is an increased incidence when depolarising muscle relaxants are used in combination with inhaled volatile gases.23 It is more prevalent in children, with approximately 50% of cases occurring before the age of 15. Table 7 shows the clinical features. Susceptibility tests (mainly the in vitro contraction test) may be diagnostic and can be applied to family members after an affected individual has been identified. The in vitro contraction test requires a large fresh muscle biopsy after which either halothane or caffeine may be applied and the maximal contraction measured.

Disordered muscle calcium regulation is now known to underlie the pathophysiology of malignant hyperthermia syndromes. A trigger (for example, general anaesthesia) leads to excessive activation of the ryanodine receptor calcium release channel and thus calcium is released from sarcoplasmic reticulum stores. Calcium reuptake from the cytoplasm may also be impaired. The increased cytoplasmic calcium leads to excessive muscle contraction, hypermetabolism, rhabdomyolysis, and fever.26 Dantrolene inhibits release of calcium from sarcoplasmic reticulum27 and early administration has reduced the mortality rate from 70% to approximately 10%. Mutations in the ryanodine receptor (RYR1) on chromosome 19q1326 are found in 50% of families with

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**Table 6 Clinical features of Andersen’s syndrome**

<table>
<thead>
<tr>
<th>System</th>
<th>Features</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cardiac</td>
<td>Prolonged QT interval is common (early sign)</td>
</tr>
<tr>
<td></td>
<td>Ventricular arrhythmia (may segregate in females)</td>
</tr>
<tr>
<td></td>
<td>Bigeminy, bidirectional ventricular tachycardia, complete heart block</td>
</tr>
<tr>
<td></td>
<td>Symptoms: syncope, sudden death</td>
</tr>
<tr>
<td></td>
<td>Treatment: poor response to classical therapy but amiodarone may have a role</td>
</tr>
<tr>
<td>Skeletal muscle</td>
<td>Episodic weakness (may segregate in males)</td>
</tr>
<tr>
<td></td>
<td>Age of onset 2 to 18 years; duration 1 hour to days; precipitants K+, exercise, or none</td>
</tr>
<tr>
<td></td>
<td>There is no associated myotonia</td>
</tr>
<tr>
<td></td>
<td>Occasionally there is permanent weakness which may be proximal or distal</td>
</tr>
<tr>
<td>Face</td>
<td>Hypertelorism, mandibular hypoplasia, low set ears, broad forehead, malar hypoplasia</td>
</tr>
<tr>
<td></td>
<td>The severity of dysmorphic features does not correlate with cardiac or skeletal muscle involvement</td>
</tr>
<tr>
<td></td>
<td>Dysmorphism may be mild and overlooked unless specifically considered</td>
</tr>
<tr>
<td>Other</td>
<td>Hypoplastic kidney, cardiac malformations (for example, semilunar valve abnormalities)</td>
</tr>
</tbody>
</table>

**Table 7 Clinical features of malignant hyperthermia**

<table>
<thead>
<tr>
<th>System</th>
<th>Features</th>
</tr>
</thead>
<tbody>
<tr>
<td>Skeletal muscle</td>
<td>Rigidity and weakness</td>
</tr>
<tr>
<td></td>
<td>Rhabdomyolysis</td>
</tr>
<tr>
<td></td>
<td>Muscle spasms especially affecting masseter, but can be generalised</td>
</tr>
<tr>
<td></td>
<td>Myalgia</td>
</tr>
<tr>
<td>Autonomic</td>
<td>Sym pathetic overactivity</td>
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<tr>
<td></td>
<td>Hyperventilation</td>
</tr>
<tr>
<td></td>
<td>Tachycardia</td>
</tr>
<tr>
<td></td>
<td>Haemodynamic instability</td>
</tr>
<tr>
<td></td>
<td>Cardiac arrhythmia</td>
</tr>
<tr>
<td>General</td>
<td>Fever (may be a late sign)</td>
</tr>
<tr>
<td></td>
<td>Cyanosis</td>
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<tr>
<td>Laboratory</td>
<td>Increased oxygen consumption</td>
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<tr>
<td></td>
<td>Hypercapnia</td>
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<tr>
<td></td>
<td>Lactic acidosis</td>
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<td></td>
<td>Raised creatine kinase</td>
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<tr>
<td></td>
<td>Hyperkalaemia</td>
</tr>
<tr>
<td>Triggers</td>
<td>Full episodes: general anaesthesia (inhaled anesthetics—Isoflurane, desflurane, enflurane, sevoflurane, methoxyflurane and halothane), suxamethonium</td>
</tr>
<tr>
<td></td>
<td>Milder malignant hyperthermia: exercise in hot conditions, neuroleptic drugs, alcohol, infections</td>
</tr>
<tr>
<td>Treatment</td>
<td>Dantrolene 2 mg/kg intravenously every 5 minutes to a total of 10 mg/kg</td>
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<tr>
<td></td>
<td>Hyperventilation with supplemental oxygen</td>
</tr>
<tr>
<td></td>
<td>Sodium bicarbonate</td>
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<tr>
<td></td>
<td>Active cooling</td>
</tr>
<tr>
<td></td>
<td>Discontinue anaesthesia</td>
</tr>
<tr>
<td></td>
<td>Maintain urine output over 2 ml/kg/hour</td>
</tr>
<tr>
<td></td>
<td>Avoid calcium, calcium antagonists, β-blockers</td>
</tr>
</tbody>
</table>
malignant hyperthermia and 20% of all patients with malignant hyperthermia. So far over 30 mutations have been identified, most are missense and 50% lie between exons 39 to 46. However, there is genetic heterogeneity, with at least five dominantly inherited susceptibility loci identified. These include (1) the sodium channel α-subunit (SCN4A), allelic with HyperKPP, (2) the skeletal muscle voltage dependent L-type calcium channel (dihydropyridine receptor) α2δ-subunit (CACNL2A1), (3) the L-type calcium channel (dihydropyridine receptor) α2δ-subunit (CACNA1S), allelic with HypoKPP, and (4) unknown genes located on chromosome 3q13.1 (MHS4) and chromosome 5p (MHS6).

There are also several other primary muscle disorders with an associated susceptibility to an malignant hyperthermia-like reaction. The term malignant hyperthermia-like is used to indicate that these patients may develop the symptom complex very similar to that described in malignant hyperthermia but in these cases there is not a primary disturbance of muscle calcium handling—that is, they do not have RyR mutations. It is suspected that they have a tendency to a disturbance of calcium handling which seems to be secondary to their primary disease. These include myotonia congenita, periodic paralysis, myotonic dystrophy type I, Duchenne and Becker muscular dystrophy, mitochondrial disorders, carnitine palmitoyl-transferase deficiency, and Brody's myopathy. Caution in relation to anaesthesia is therefore advised in patients in all these groups. We advise all patients with these disorders to ensure their anaesthetist and surgeon are aware of the potential for a malignant hyperthermia-like reaction. Similar anaesthetic precautions can then be taken for this patient group.

Central core disease (malignant hyperthermia syndrome 1)

This is a congenital myopathy with susceptibility to malignant hyperthermia. The clinical features include a non-progressive myopathy with facial and proximal weakness and hypotonia. Occasionally muscle cramps after exercise are seen. More than a quarter of patients with central core disease have a tendency to malignant hyperthermia, however around 40% of cases at risk for malignant hyperthermia are asymptomatic. In such cases adequate precautions before anaesthesia are impossible. Central core disease is characterised pathologically by the presence of central core lesion throughout the length of type I muscle fibres. Missense mutations in the skeletal muscle ryanodine receptor gene (RYR1) have been identified in some families with central core disease.

Investigation after an episode of malignant hyperthermia

A full blown case of malignant hyperthermia is usually a dramatic clinical presentation familiar to anaesthetists. Confirmation of susceptibility to recurrent attacks after such a full attack can be achieved in specialist centres by the in vitro contraction test in combination with genetic testing.

It is very important to screen family members related to any individual who has had such a full blown attack. This must include a careful history, including symptoms of muscle disease—for example, cramps, myalgia, fatigue, myoglobinuria—family history of anaesthetic complications, and measurement of baseline creatine kinase and urine examination for myoglobinuria. Referral to a centre where muscle biopsy, in vitro contraction test, and genetic testing are available is usually required to be certain about status of potentially at risk family members. In the interim, all at risk relatives should be warned of a possible increased risk of malignant hyperthermia under general anaesthesia and be advised to inform their surgeon and anaesthetist. Non-urgent surgery should be postponed until the diagnosis is clarified. However, general anaesthesia can be safely administered if the anaesthetist is aware of the risk and the proper precautions are instituted. Patients should be advised to wear a Medic-alert bracelet.

Neuromuscular junction

Congenital myasthenic syndromes

There are several rare congenital myasthenic syndromes due to defects in the key processes that underlie efficient neuromuscular junction transmission. The commonest are mutations in the subunits of the postsynaptic acetylcholine receptor. These myasthenic syndromes may therefore be considered to be genetic ligand gated channelopathies. A detailed summary is beyond the scope of this review, but interested readers are directed to the review by Engel et al.

Central nervous system

In the last few years an increasing number of genetic CNS channelopathies have been described. Although the starting point for many of these studies were individual families with rare syndromes (for example, familial hemiplegic migraine or benign familial neonatal convulsions), there is increasing evidence that the discoveries made will be relevant to common neurological diseases such as migraine and epilepsy. Perhaps the best evidence that ion channel dysfunction is important in common neurological disease is the recent evidence in epilepsy described below. It has been shown that a particular epilepsy phenotype know as “generalised epilepsy with febrile seizures” is more common than previously realised and that it frequently associates with mutations in brain ion channel genes.

Familial hemiplegic migraine

Familial hemiplegic migraine is a form of migraine with aura which is inherited in an autosomal dominant manner. Patients experience typical migraine headaches but in addition there are paroxysmal neurological symptoms of aura including hemianopia, hemisensory loss, and dysphasia. Hemiparesis occurs with at least one other symptom during familial hemiplegic migraine aura; the weakness can be prolonged and may outlast the associated migrainous headache by days. Coma has also been described with severe attacks. Persistent attention deficits and memory loss can last weeks to months. Triggers include emotion or head injury. The age at onset for familial hemiplegic migraine is often earlier than typical migraine, frequently beginning in the first or second decade. The number of attacks tends to decrease with age. About 20% of families have cerebellar signs ranging from nystagmus to progressive, usually late onset cerebellar ataxia. Genetic studies have established that many cases of familial hemiplegic migraine are caused by missense mutations in the P/Q-type voltage gated calcium channel gene, CACNA1A. The presynaptic location of this calcium channel allows it to function as a key controller and modulator of the release of both excitatory and inhibitory neurotransmitters throughout the CNS. It is suspected that a disturbance in this control is important in the genesis of familial hemiplegic migraine.

Episodic ataxia

The episodic ataxias are rare autosomal dominant CNS disorders in which the main clinical features are episodes of profound cerebellar ataxia. The clinical features of episodic ataxias type 1 and 2 are summarised in table 8. In patients with episodic ataxia type 1 experiences very brief episodes of sudden onset ataxia that may be precipitated by sudden movement or emotion. There may be multiple attacks in a day. Cerebellar function is normal between attacks but there is persistent myokymia or neuromyotonia of...
Episodic ataxia type 2.40
voltage gated calcium channel, CACNA1A, cause episodic
vented with acetazolamide treatment. Mutations in the
clinical features.
the neuromuscular junction, hence the combination of
terminal region of the gene).41 45
an expansion of a CAG repeat normally present in the C
coding for C terminal of the calcium channel protein (that is,
trinucleotide repeat expansion in the region of the gene
progressive pure cerebellar ataxia and typically associates with a
spinocerebellar ataxia type 6 (SCA6) has also been shown to
ataxia type 2 described above a third disorder known as
skeletal muscles. Clinically, this may be observed as fine
twitching movements around the eyes or in the limbs. This
often produces involuntary fine side-to-side movements
of the fingers in the outstretched hands. The myokymia/
neuromyotonia is clinically and electrophysiologically indis-
tinguishable from that seen in the autoimmune potassium
channelopathy Isaac’s syndrome (see below). Episodic ataxia
type 1 is caused by mutations in the potassium channel gene
KCNA1,42–44 which is expressed both in the cerebellum and at
the neuromuscular junction, hence the combination of
clinical features.

Episodic ataxia type 2 is characterised by prolonged attacks
of cerebellar ataxia. The patient is profoundly ataxic and
often has a prominent headache and a feeling of vertigo and
nausea. It is probable that many patients previously labelled
as having basilar migraine in fact have episodic ataxia type 2.
Attacks are precipitated most commonly by emotion and
occasionally by intercurrent illness. The attack frequency
decreases with age but some patients develop a progressive
cerebellar atrophy.45 Attacks are often successfully pre-
vented with acetazolamide treatment. Mutations in the
voltage gated calcium channel, CACNA1A, cause episodic
ataxia type 2.40

Calcium channel allelic disorders familial
hemiplegic migraine, episodic ataxia type 2,
spinocerebellar ataxia type 6: molecular
mechanisms
In addition to familial hemiplegic migraine and episodic
ataxia type 2 described above a third disorder known as
spinocerebellar ataxia type 6 (SCA6) has also been shown to
associate with a mutation in the same calcium channel gene
CACNA1A. SCA6 is a late onset autosomal dominant pro-
gressive pure cerebellar ataxia and typically associates with a
trinucleotide repeat expansion in the region of the gene
coding for C terminal of the calcium channel protein (that is,
an expansion of a CAG repeat normally present in the C
terminal region of the gene).41 45

Table 8 Clinical features of episodic ataxia

<table>
<thead>
<tr>
<th></th>
<th>Episodic ataxia type 1</th>
<th>Episodic ataxia type 2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mode of inheritance</td>
<td>Autosomal dominant</td>
<td>Autosomal dominant</td>
</tr>
<tr>
<td>Age of onset</td>
<td>Second decade</td>
<td>Early childhood to teens</td>
</tr>
<tr>
<td>Clinical features</td>
<td>Ataxia, Dizziness without vertigo, Visual blurring, No nystagmus</td>
<td>Ataxia, truncal instability which may persist between attacks, dysarthria, nystagmus, Associated with vertigo, nausea, vomiting, and headache</td>
</tr>
<tr>
<td>Exacerbating factors</td>
<td>Abrupt postural change, emotion, starle, vestibular stimulation</td>
<td>Physical or emotional stress</td>
</tr>
<tr>
<td>Duration of attack</td>
<td>Brief, attacks last minutes</td>
<td>Attacks often last 30 minutes to &gt;24 hours</td>
</tr>
<tr>
<td>Additional features</td>
<td>Neuromyotonia (continuous spontaneous muscle fibre activity) or myokymia occur during and between episodes of ataxia Some patients have hyperhidrosis and seizures</td>
<td>Downbeating gaze evoked nystagmus in all directions between episodes. Impaired vestibulo-ocular reflex, OKN and smooth pursuits. Some patients develop progressive cerebellar atrophy</td>
</tr>
<tr>
<td>Treatment</td>
<td>Phenytoin, carbamazepine, not acetazolamide</td>
<td>Acetazolamide</td>
</tr>
<tr>
<td>Ion channel gene</td>
<td>Potassium: KCNA142–44</td>
<td>Calcium: CACNA1A,46 allelic with FHM and SCA6</td>
</tr>
</tbody>
</table>

It is therefore evident that familial hemiplegic migraine,
episodic ataxia type 2, and SCA6 are different clinical
phenotypes caused by mutations in the same gene—that is,
they represent allelic disorders. Although there are some
genotype phenotype correlations, there is also overlap.
Furthermore, the precise molecular mechanisms underlying
the different phenotypes are not elucidated. Some general,
although not absolute, observations are emerging and are
briefly outlined below.

Familial hemiplegic migraine most frequently associates
with missense mutations in CACNA1A. Expression studies
have shown various consequences of these missense muta-
tions on channel function but broadly speaking an alteration
in channel kinetics is observed. Both an increase and a
decrease in channel kinetics have been reported making it
difficult to produce a unifying hypothesis for the genesis of
the migraine attacks.41

Episodic ataxia type 2 most commonly associates with
point mutations in CACNA1A which are predicted to truncate
the calcium channel protein. Expression studies have pointed
to a loss of function and haploinsufficiency as the basis of the
attacks.45

SCA6 virtually always associates with a CAG repeat
expansion in CACNA1A as described above. Unlike other
CAG repeat expansions observed in neurogenetic diseases
such as in Huntington’s disease the SCA6 expansion is
relatively stable on transmission and the phenomenon of
clinical anticipation (that is, the worsening of disease severity
as judged by earlier age at onset in succeeding generations
frequently observed in Huntington’s disease) is not observed
in SCA6. Evidence has been produced that the SCA6
expansion may not only reduce calcium channel function
but, as reported in other neurological trinucleotide repeat
diseases, may result in abnormal aggregation of calcium
channel protein harbouring expanded glutamine tracts coded
for by the CAG repeat.

Exceptions to these general observations apply both at a
clinical as well as expression level. For example, there are
missense mutations described which cause a pure progressive ataxia without familial hemiplegic migraine or episodic ataxia type 2 features. Furthermore an episodic ataxia type 2 phenotype has been reported in patients harbouring the SC1A6 expansion. Further study of molecular mechanisms is clearly required.

**Hypercereplexia**

Hypercereplexia is characterised by onset at birth with hypertonia that disappears in sleep, exaggerated startle response, and strong brainstem reflexes (especially the head retraction reflex). The exaggerated startle reaction occurring after sudden, unexpected acoustic or tactile stimuli persists into adulthood and is associated with involuntary myoclonus (occasionally resulting in falls) and marked nocturnal myoclonic jerks. Continuous and occasionally fatal muscular rigidity is also a feature. Electromyography shows continuous motor unit activity. The GLRA1 gene (encoding the glycine receptor γ1-subunit, a ligand gated ion channel) was the first gene for a neurotransmitter receptor in the CNS to be identified as the site of mutation in a human disorder.44 Cases are usually autosomal dominant, but some recessive pedigrees have been reported.

**Andermann’s syndrome**

Andermann’s syndrome is an autosomal recessive hereditary motor and sensory neuropathy with agenesis of the corpus callosum. This is found at high frequency (1:2100 live births) in the province of Quebec in Canada. Mutations impair function of the potassium-chloride co-transporter, KCC3 found in the brain and spinal cord encoded by SLC12A647 on chromosome 15q13–q15. There is symmetric or asymmetric involvement of the cranial nerves with ptosis, facial weakness, ophthalmoplegia (reduced upgaze), and optic atrophy. Motor neuropathy presents early with hypotonia and severe progressive global weakness; affected patients rarely walk independently. Sensory loss is manifest as areflexia and tremor. Involvement of the CNS is seen as mental retardation, seizures, and atypical psychosis with onset in the teens. Dysmorphic features include long facies, hypertelorism, brachycephaly, high arched palate, syndactyly of the second and third toes, and overriding the first toe. Scoliosis may lead to a restrictive lung defect. There is partial or complete agenesis of the corpus callosum due to a defect in axon migration across the midline.

**Inherited epilepsy syndromes**

There are several inherited epilepsy syndromes which are genetic channelopathies. Here we discuss four of these.

**Benign familial neonatal convulsion**

This is an autosomal dominant disorder characterised by brief generalised seizures that clear spontaneously after the age of 6 weeks, with no neuropsychological morbidity. Knowledge of this disorder can prevent needless and potentially harmful anticonvulsant therapy. It is now established that benign familial neonatal convulsion is a CNS potassium channelopathy. Highly penetrant mutations in the KCNQ2 gene on chromosome 20 and KCNQ3 gene on chromosome 8 have been found in pedigrees with benign familial neonatal convulsion.

**Generalised epilepsy with febrile seizures plus syndrome**

One of the most important genetic epilepsy discoveries in recent years has been the identification of the generalised epilepsy with febrile seizures plus syndrome phenotype.45 This is a pleomorphic familial epilepsy syndrome that may include febrile and afebrile generalised tonic-clonic, absence, myoclonic, and atonic seizures. Any combination of seizures may be observed in a given family. There are now a significant number of such families described worldwide. Generalised epilepsy with febrile seizures plus syndrome has been found to be genetically heterogeneous, but in all cases disturbed ion channel function has been found to be the basis of the disease. Mutations have been described in the voltage gated sodium channel β1-subunit gene (SCN1B)45 46; voltage gated sodium channel γ1-subunit gene (SCN1A)47; GABA receptor γ2-subunit gene (GABRG2)48 49; and the voltage gated sodium channel type II γ2-subunit gene (SCN2A).50 It has been shown that increased neuronal excitability due to the mutant channels is predicted to lead to epileptogenesis. Since the possible epilepsy phenotypes that may occur in these families with normal neuroimaging are indistinguishable from many common epilepsy phenotypes, many workers are now undertaking studies to establish if variations in these channel genes are important in determining susceptibility to common forms of epilepsy.

**Autosomal dominant nocturnal frontal lobe epilepsy**

This is characterised by focal onset frontal lobe seizures, almost exclusive occurrence during drowsiness or sleep, and variable severity of symptoms in family members. Milder cases are often undiagnosed or misdiagnosed as nightmares, parasomnias, or functional disorders. Neuroimaging is normal and treatment with carbamazepine is dramatically effective. Although recognition of this syndrome is important for appropriate therapy and genetic counselling, underestimation of cases is likely. The clinical delineation of this as a separate disorder allowed genetic analysis to be performed.46 Mutations have been found in the nicotinic acetylcholine receptor γ2-subunit, CHRNA448 and β3-subunit, CHRN8.48 This is a brain ligand gated ion channel that is mainly presynaptic in location and has a role in controlling neurotransmitter release. Alterations in the balance of excitatory and inhibitory transmitters are suggested to be important in the genesis of seizures.

**Childhood absence epilepsy**

Most recently, studies in a large Chinese cohort with childhood absence epilepsy found mutations in the brain T-type calcium channel CACNA1H.49 Functional analysis has shown that two of the mutations allow increased calcium influx during physiological activation and another results in channel opening at more hyperpolarised potentials, which may underlie the propensity for seizures.50 However, a recent study has failed to replicate these findings in mixed idiopathic generalised epilepsy pedigrees.51

**Autoimmune Channelopathies**

For comparison to selected genetic channelopathies, see table 9.

**Neuromuscular junction**

Myasthenia gravis is the archetypal autoimmune channelopathy affecting neuromuscular transmission, via the acetylcholine receptor, a ligand gated ion channel. There are many excellent descriptions of this disorder in standard textbooks and we will not consider it further here.

**Lambert-Eaton myasthenic syndrome**

Lambert-Eaton myasthenic syndrome (LEMS) is a paraneoplastic disorder in which patients produce antibodies directed against the presynaptic voltage gated P/Q-type calcium channel α1A-subunit. Typically, patients experience proximal weakness often in combination with autonomic symptoms. A common finding is absent or diminished tendon reflexes which reappear after brief maximal voluntary contraction or...
repeated tendon percussion (post-tetanic potentiation). Weakness, when present, almost universally affects the lower limbs. The upper limbs are frequently affected with involvement of bulbar and respiratory musculature less often. Weakness may be improved with brief exercise and may worsen with sustained exercise, heat, or fever. Fatigability is present in a third of cases. Some patients complain of myalgia. In contrast to myasthenia gravis the extraocular muscles are infrequently involved. There may also be an associated distal, symmetric sensory neuropathy. As in myasthenia gravis, LEMS may occasionally be exacerbated by drugs—for example, neuromuscular blocking agents, antibiotics (aminoglycosides, fluoroquinolones), magnesium, Ca\(^{2+}\) channel blockers, and iodinated intravenous contrast agents. However, in contrast to myasthenia gravis, LEMS never begins with ocular weakness and usually has more marked weakness in legs than in the arms.

Antibody-mediated reduction in the number of presynaptic calcium channels at the nerve terminal leading to reduced acetylcholine release underlies the pathogenesis of LEMS. IgG antibodies against the P/Q-type calcium channel \(\alpha_{1A}\) subunit (which is mutated in familial hemiplegic migraine, episodic ataxia type 2, and SCA6) are present in over 85% of cases. Repetitive nerve stimulation shows an increment after rapid or sustained muscle contraction that is prolonged by cooling muscles. Small cell lung carcinoma (SCLC) is the most frequently associated neoplasm. Indeed, up to 3% of patients with SCLC have LEMS. Other associations include lymphoproliferative disorders—for example, reticulum cell sarcoma, T-cell leukaemia, lymphoma, and Castleman’s disease. The onset of LEMS is usually six months to five years before any neoplasm is detected. However, one third of cases occur in the absence of malignancy at diagnosis and do not seem to develop a tumour even over long term follow up. Such cases may represent primary autoimmune non-paraneoplastic disorders. Treatment is often beneficial, the mainstay being 3,4-diaminopyridine. There may be an associated cerebellar syndrome (see below).

**Peripheral nervous system**

**Isaac’s syndrome**

Acquired neuromyotonia (Isaac’s syndrome) manifests as muscle cramps, slow relaxation of muscles after contraction (pseudomyotonia) and hyperhidrosis. Electromyography shows myokymic and neuromyotonic discharges (repetitive firing at rates of 5–150 Hz and 150–300 Hz respectively). This spontaneous muscle activity is driven by abnormal firing of peripheral nerves. Antibodies to voltage gated potassium channels have been shown to induce hyperexcitability of the nerve by suppressing the outward potassium current. It is evident that Isaac’s syndrome is the autoimmune counterpart to genetically determined neuromyotonia. In both situations the same potassium channel is dysfunctional. In Isaac’s syndrome this is induced by autoimmune attack, in contrast, in genetic neuromyotonia there is a mutation in the gene for the same potassium channel—that is, the KCNA1 gene.

**Central nervous system**

**Paraneoplastic cerebellar degeneration**

Paraneoplastic cerebellar degeneration (PCD) usually presents as a subacute cerebellar syndrome which progresses over weeks to months. There are multiple antineuronal antibodies associated with PCD, the most common of which are anti-Yo in breast and ovarian malignancies and anti-Hu in SCLC. Some patients with LEMS have also been noted to have cerebellar ataxia, implicating the P/Q-type calcium channel in pathogenesis. This channel is expressed not only by presynaptic PNS nerve terminals but also those in cerebellar Purkinje and granule cells. Postmortem findings in these patients show Purkinje cell loss and cerebellar cortical gliosis. In one study, 9% of LEMS patients had coexistent PCD and high titres of anti-P/Q-type voltage gated calcium channel antibodies (the same channel affected in SCLC). In another study of PCD, 41% of patients were found to have anti-P/Q-type voltage gated calcium channel antibodies with accompanying cerebrospinal fluid antibodies. This paraneoplastic syndrome is most frequently associated with SCLC.

PCD associated with the calcium channel antibodies described may therefore be regarded as the immunological counterpart for episodic ataxia type 2 which associates with point mutations in the same calcium channel CACNA1A. However, from a clinical viewpoint PCD is a much more aggressive, rapidly progressive cerebellar ataxia than episodic ataxia type 2. In PCD we envisage there is progressive destruction and loss of these calcium channels which may account for the clinical severity observed.

**Morvan’s syndrome**

Limbic encephalitis is a paraneoplastic syndrome associated with SCLC and rarely with other tumour types. The symptoms consist of psychiatric involvement (personality changes, hallucinations, and insomnia), seizures, short term memory loss, and confusion. There is usually hyperintense T2-weighted signal change in the hippocampi and amygdala. The majority of patients with limbic encephalitis have
associated anti-Hu antibodies. Morvan’s syndrome is the association of limbic encephalitis with neuromyotonia, hyperhidrosis, and polyneuropathy. Antibodies to voltage gated potassium channels have been found in the sera and cerebrospinal fluid of these patients. Reducing the antibody titres with plasma exchange has led to symptomatic improvement.

CONCLUSIONS
The neurological channelopathies are an important and expanding area within neurology. It is evident that the PNS and CNS may be affected in isolation or in combination. In addition, it has become clear that either genetic or autoimmune insults to the relevant channel may underlie disease pathogenesis. Indeed, it may well transpire that for each genetic channelopathy there will be its autoimmune counterpart, for examples, see table 9. To date, autoimmune channelopathies affecting the CNS are relatively uncommon but are likely to increase as further antibodies are identified. For example, the true incidence of Morvan’s syndrome is not established and it is likely to be under recognised. Furthermore, the role of antibodies directed against the CNS in the genesis of epilepsy has not been fully elucidated. For many channelopathies an accurate genetic or autoimmune diagnosis can be achieved. For muscle genetic channelopathies there is a national centre for diagnosis in the UK. Genetic diagnosis is clearly important in order to allow accurate genetic counselling in appropriate families and will often inform treatment choices. Finally, new insights into the mechanisms of epilepsy and migraine are being gained by the study of genetic channelopathies. It seems probable that genetic susceptibility to common forms of epilepsy and migraine may be determined by variation in ion channel genes, which are critical in determining neuronal excitability.

ACKNOWLEDGEMENTS
Further information regarding the UK national diagnostic service for muscle channelopathies (NSCAG service) and regarding DNA based diagnosis in neurological genetic channelopathies is available from Dr M G Hanna. The DNA diagnostic service for muscle channelopathies is supported by the Department of Health National Specialist Commissioning Agency (NSCAG), UK. Thanks to Dr Everett. Research in our laboratory is supported by the Guarantors of Brain, the Wellcome Trust, the Medical Research Council, and the Special Trustees of University College London Hospitals NHS Trust.

QUESTIONS (TRUE (T)/FALSE (F); ANSWERS AT END OF REFERENCES)
1. Consider the following statements regarding myotonia congenita:
   (A) It may be inherited in either an autosomal dominant or an autosomal recessive fashion
   (B) It affects tissues other than skeletal muscle
   (C) It is usually unresponsive to antisyntonic agents
   (D) It is caused by mutations in a voltage gated sodium channel on chromosome 17

2. Periodic paralysis:
   (A) Attacks of weakness are always accompanied by a change in serum potassium concentration
   (B) Attack frequency may be reduced by acetazolamide prophylaxis
   (C) Patients may develop permanent muscular weakness after a few years of attacks
   (D) May be caused by mutations in a ligand gated calcium channel on chromosome 1

3. Malignant hyperthermia:
   (A) May be caused by mutations in the gene encoding the ryanodine receptor of skeletal muscle
   (B) Is characterised by excessive release of calcium from the sarcoplasmic reticulum of skeletal muscle into the skeletal muscle cytoplasm
   (C) Is allelic with central core myopathy
   (D) Is precipitated by dantrolene therapy

4. Lambert-Eaton myasthenic syndrome:
   (A) Is an autoimmune channelopathy caused by antibodies against a postsynaptic voltage gated calcium channel
   (B) May be a paraneoplastic disorder frequently associated with carcinoma of the lung
   (C) Usually presents after the age of 50 years
   (D) May be associated with thymoma

5. Episodic ataxia type 2:
   (A) Is characterised by brief (1–2 minutes) attacks of ataxia
   (B) Is caused by point mutations in the voltage gated calcium channel gene CACNA1A
   (C) Is inherited in an autosomal recessive manner
   (D) Is rarely responsive to carbonic anhydrase inhibitors

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REFERENCES
Neurological channelopathies


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ANSWERS
I. A; T; B; F; C; F; D; F. 2. A; F; B; T; C; T; D; F. 3. A; T; B; T; C; T; D; F. 4. A; F; B; T; C; T; D; F. 5. A; F; B; T; C; F; D; F.